

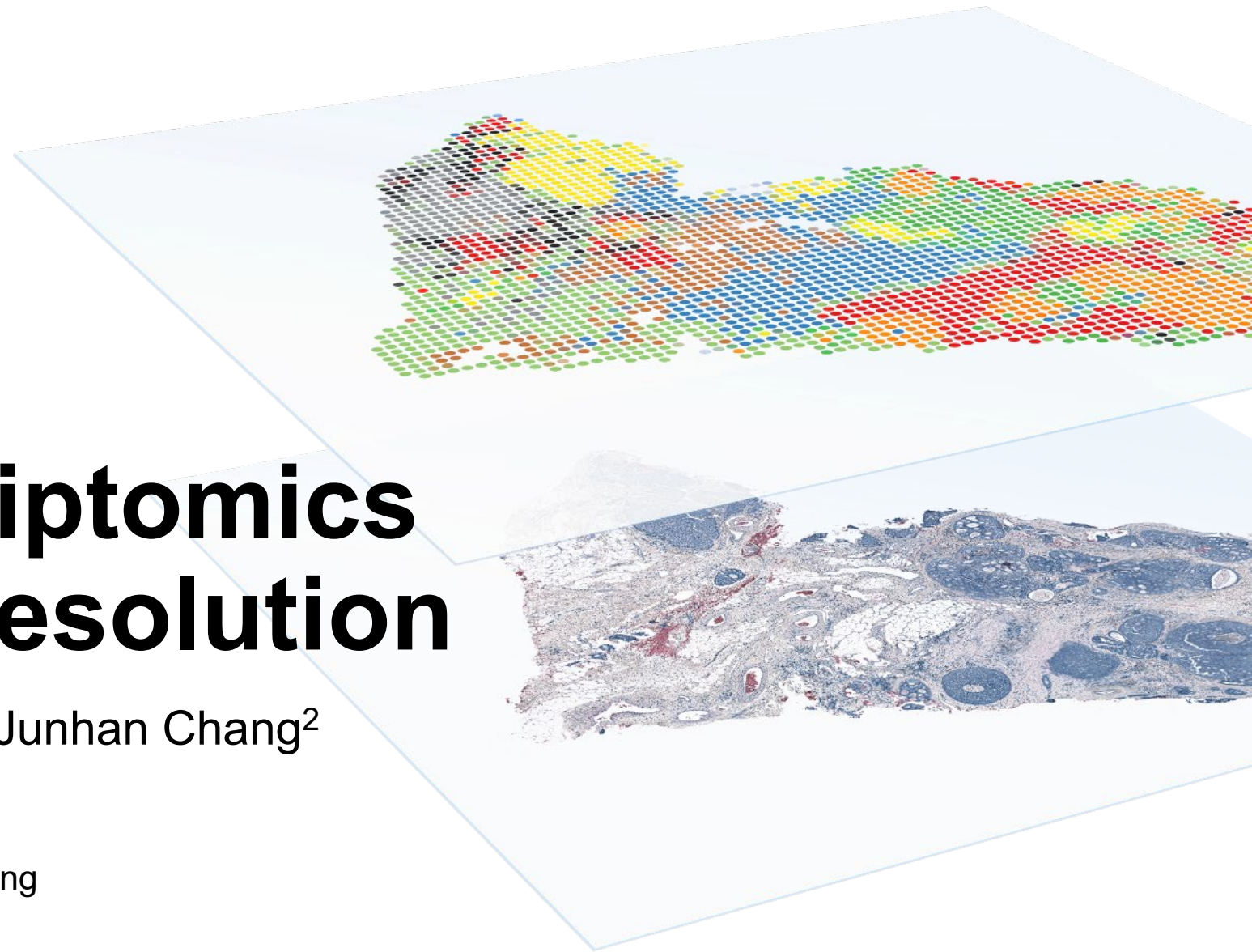
2021.05.15 UHPB Journal Club

# Spatial Transcriptomics at Single-cell Resolution

Yongcheng Jiang<sup>1</sup>, Wuji Han<sup>1</sup> and Junhan Chang<sup>2</sup>

<sup>1</sup> Integrated Science Program, Yuanpei College

<sup>2</sup> College of Chemistry and Molecular Engineering



# Honored guest of today



Prof. Letian Tao  
School of Life Sciences, PKU



Prof. Guoqiang Li  
BIOPIC, PKU

# Outline of the Journal Club

## **PART 1. Introduction**

Asp, M., Bergenstråhle, J., & Lundeberg, J. (2020). Spatially resolved transcriptomes—next generation tools for tissue exploration. *BioEssays*, 42(10), 1900221.

## **PART 2. seqFISH+**

Eng, C. H. L., Lawson, M., Zhu, Q., Dries, R., Koulena, N., Takei, Y., ... & Cai, L. (2019). Transcriptome-scale super-resolved imaging in tissues by RNA seqFISH+. *Nature*, 568(7751), 235-239.

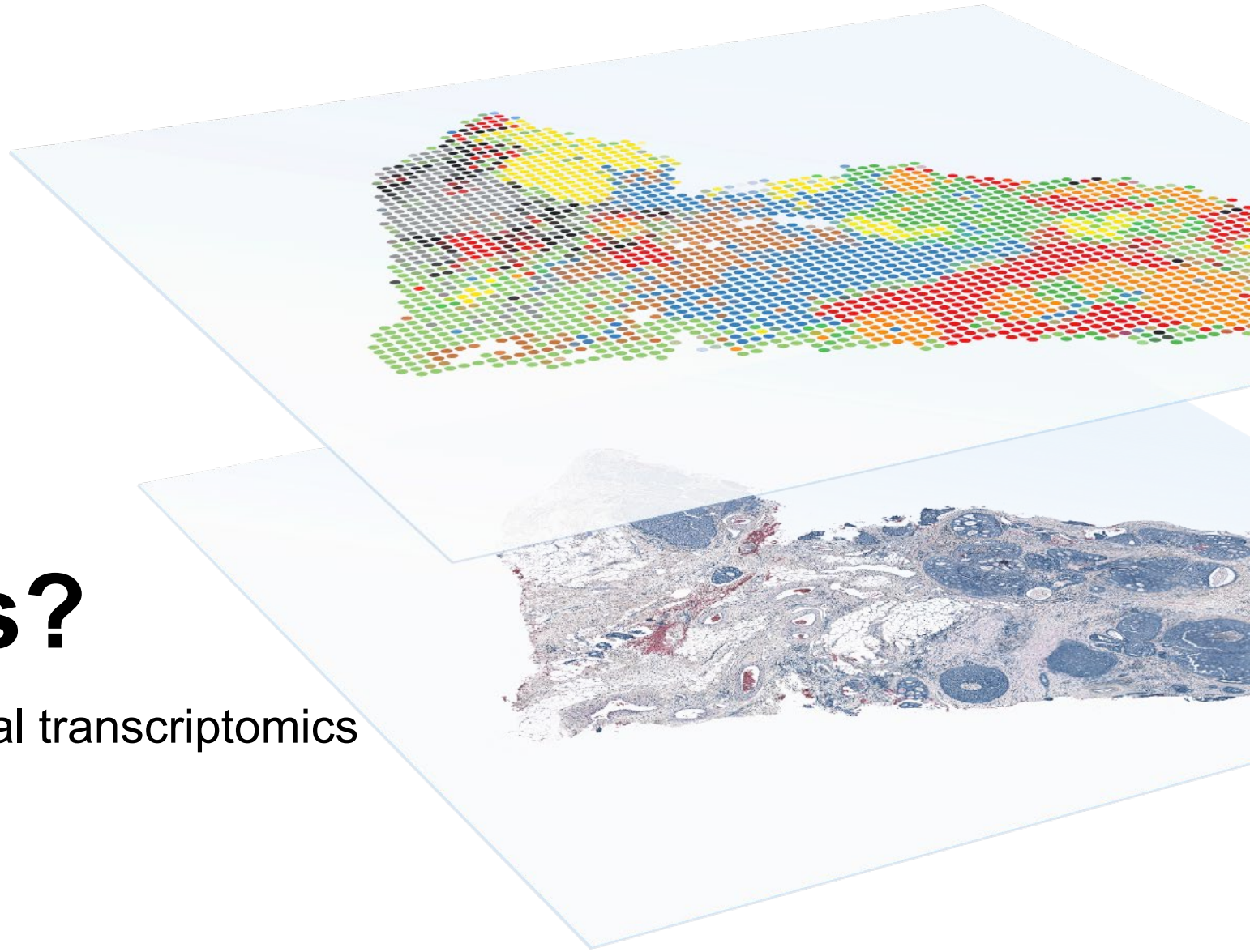
## **PART 3. CSOmap**

Ren, X., Zhong, G., Zhang, Q., Zhang, L., Sun, Y., & Zhang, Z. (2020). Reconstruction of cell spatial organization from single-cell RNA sequencing data based on ligand-receptor mediated self-assembly. *Cell research*, 30(9), 763-778.

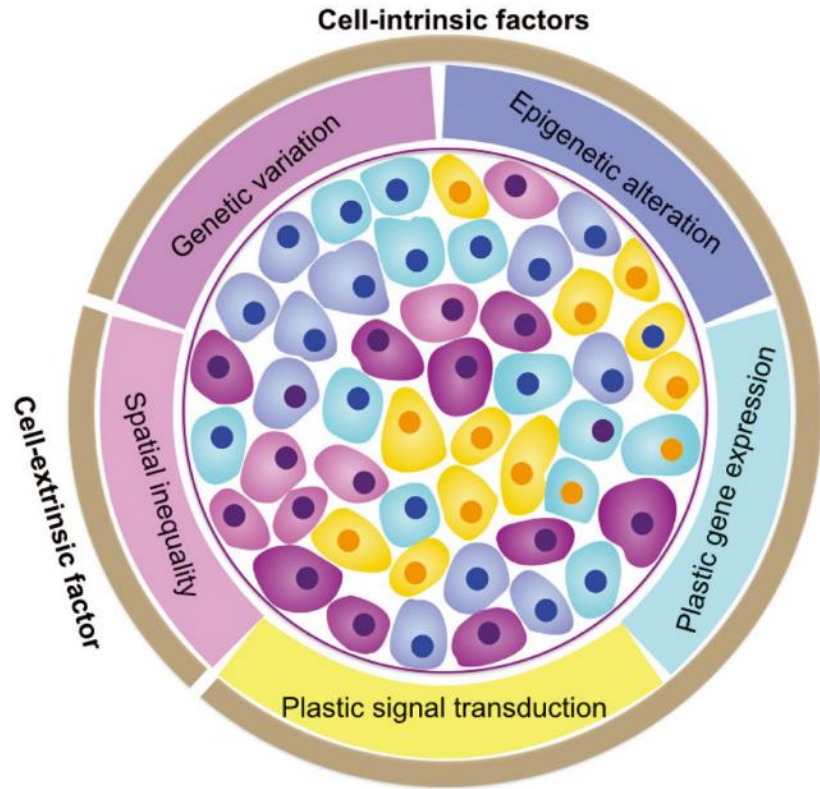
## **PART 4. Discussion**

# Why spatial transcriptomics?

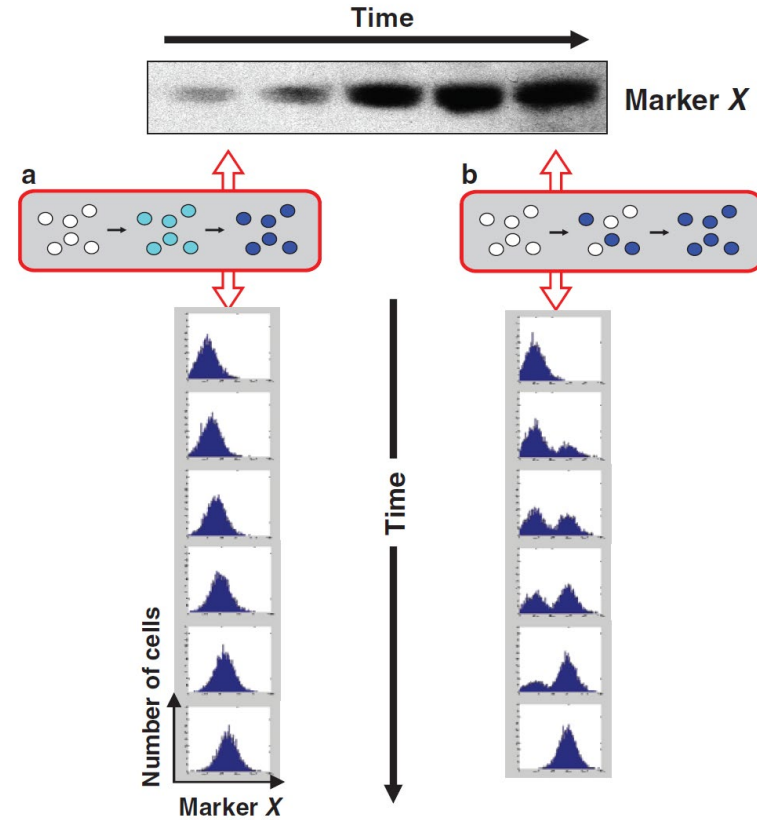
From single cell techniques to spatial transcriptomics



# Cell population shows heterogeneity

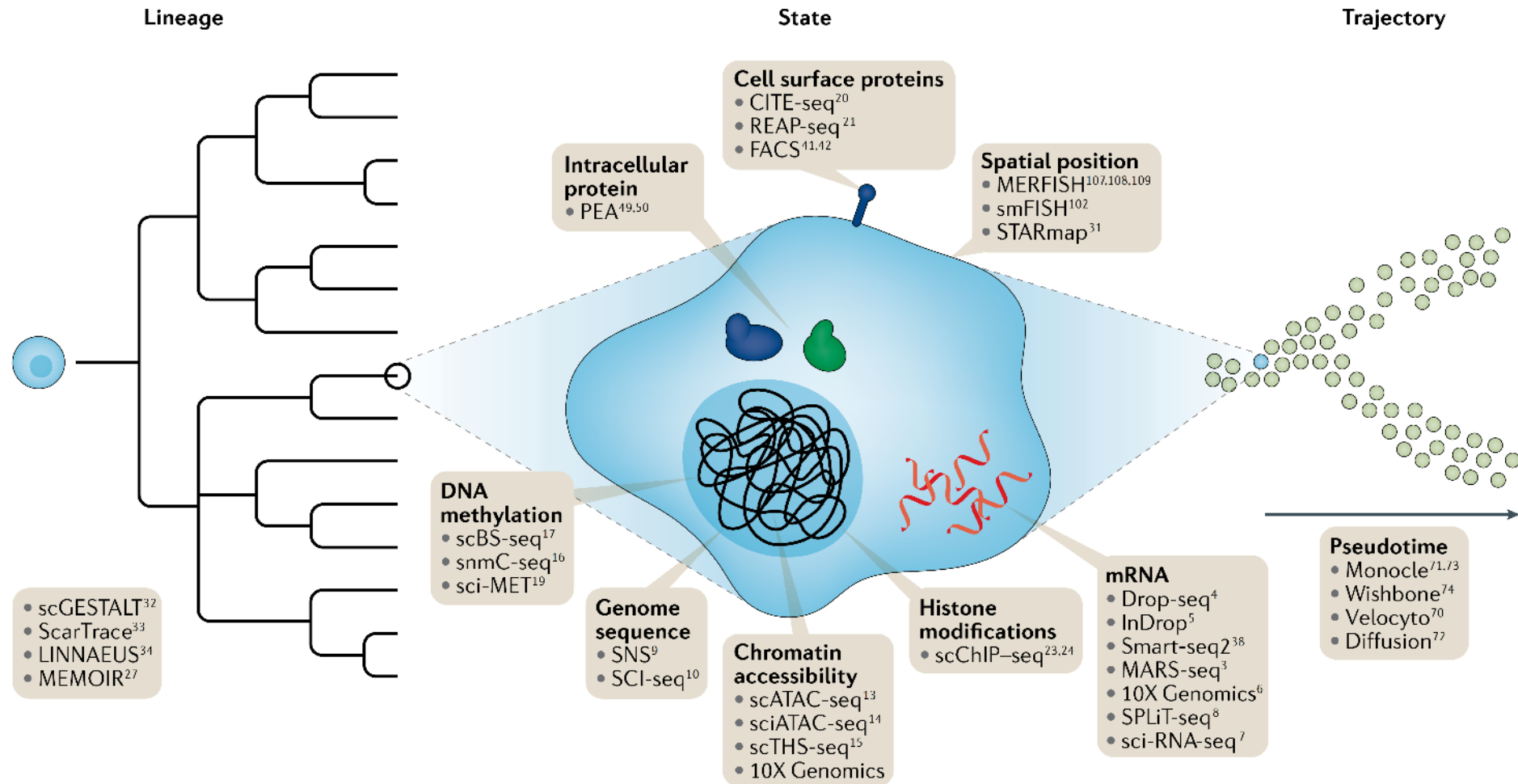


Extrinsic and intrinsic heterogeneity

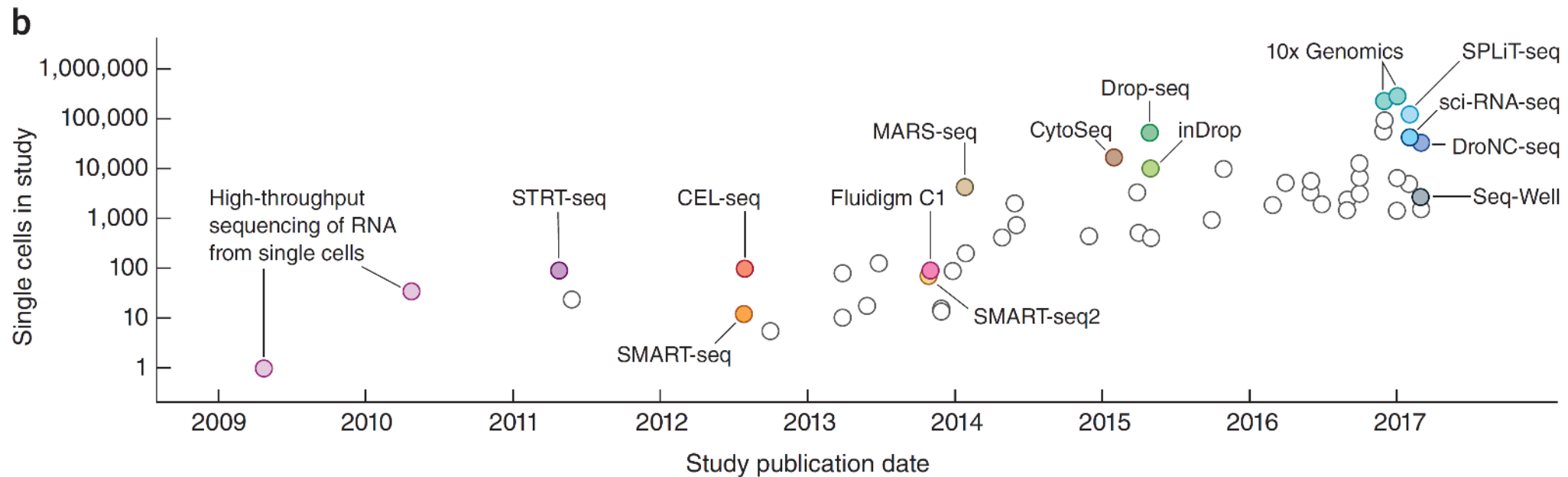
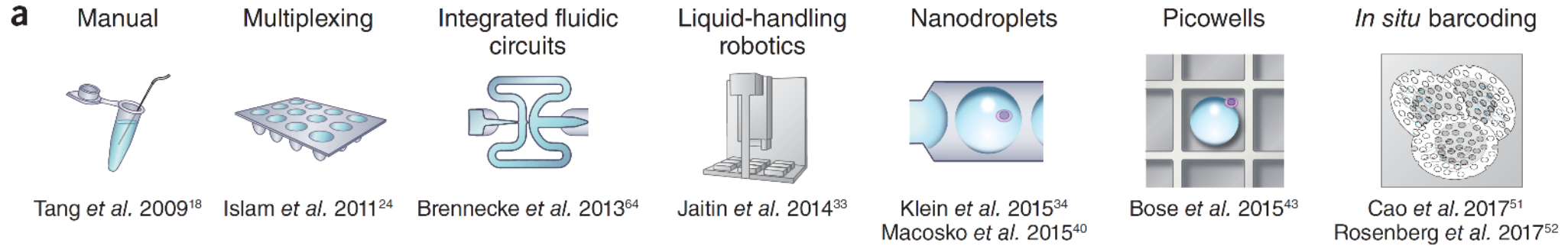


Bulk vs. single cell level data

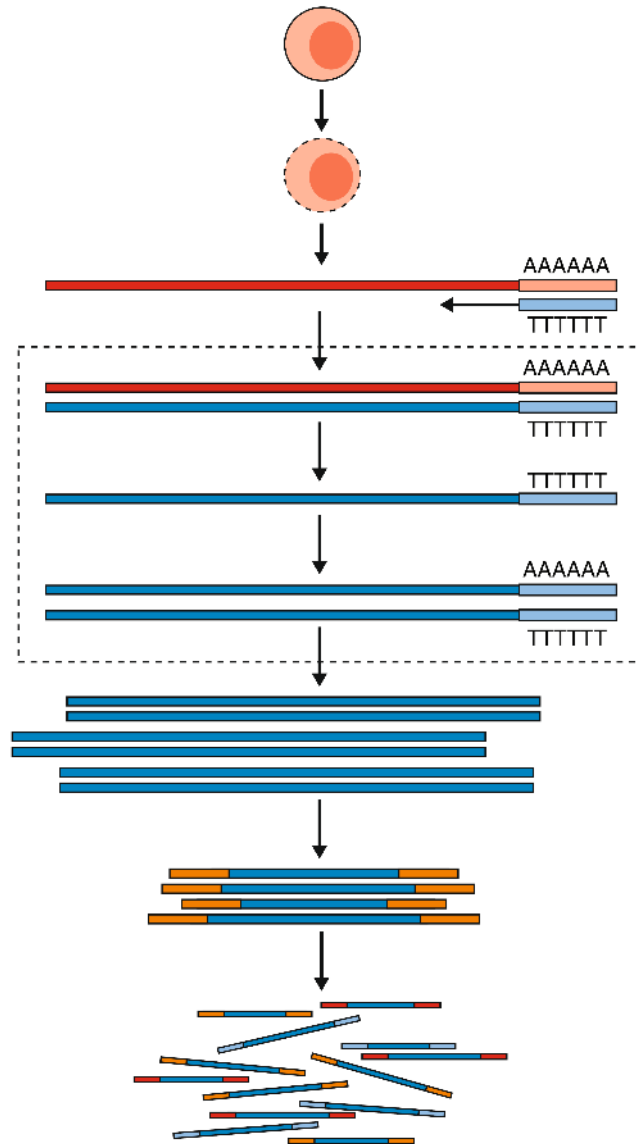
# Single-cell analysis helps reveal cellular heterogeneity



# Scaling up single-cell RNA sequencing (scRNA-seq)



# Workflow of scRNA-seq



① Isolate single cells from a tissue sample (including micro-dissection and manipulation, flow cytometric cell-sorting, microfluidic platforms, and droplet-based methods)

② Single cell lysis in a way that preserves cellular mRNA

③ mRNA molecule capture using poly(T) sequence primers that bind to mRNA poly(A) tails

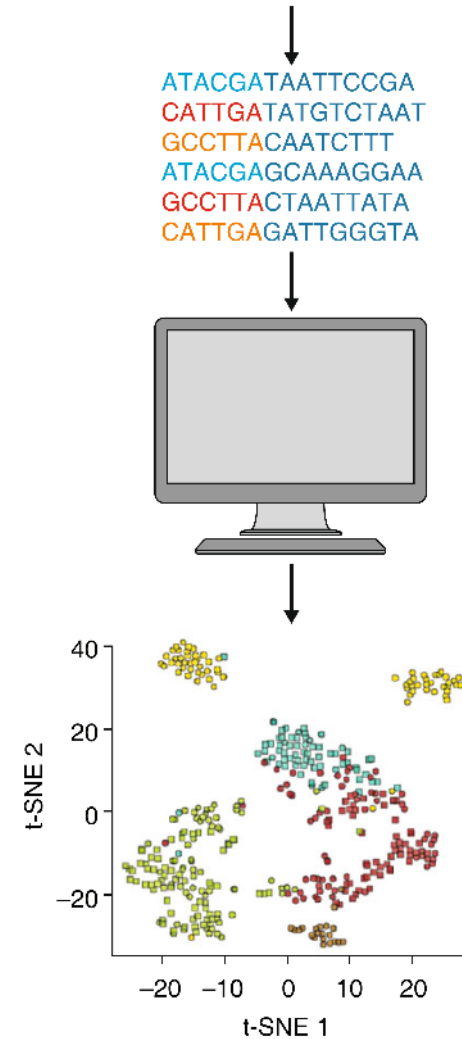
④ Convert poly(T)-primed mRNA into cDNA using reverse transcription

⑤ cDNA amplification (usually by PCR or by *in vitro* transcription)

⑥ cDNA sequencing library preparation (insert 'index' nucleotide barcodes to identify each library)

⑦ Pool cDNA sequencing libraries

Sequence libraries (via Next Generation Sequencing)

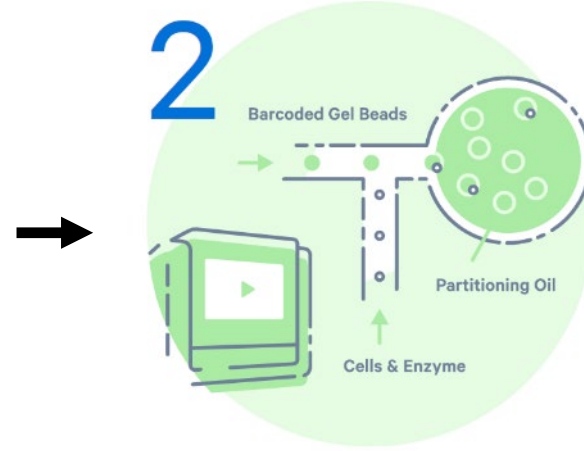




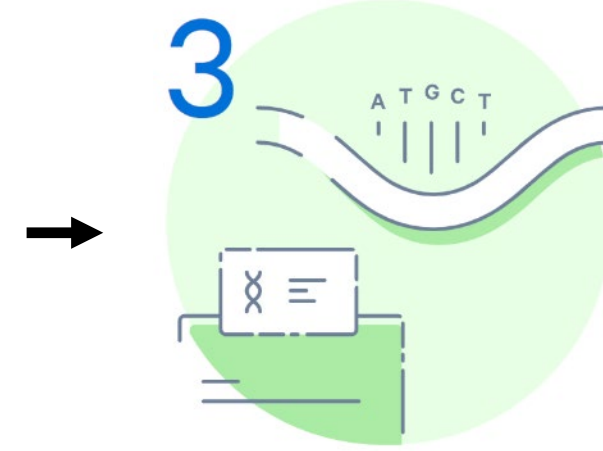
# Workflow of scRNA-seq



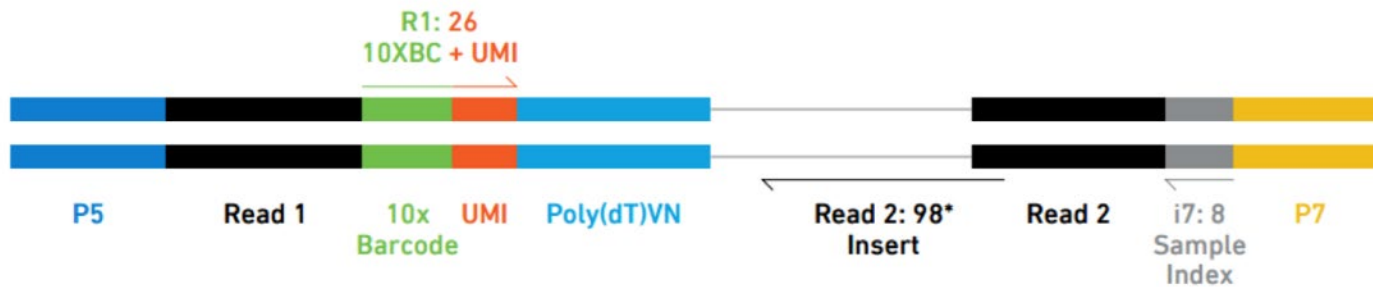
Sample preparation



Reverse transcription and library construction



Sequencing



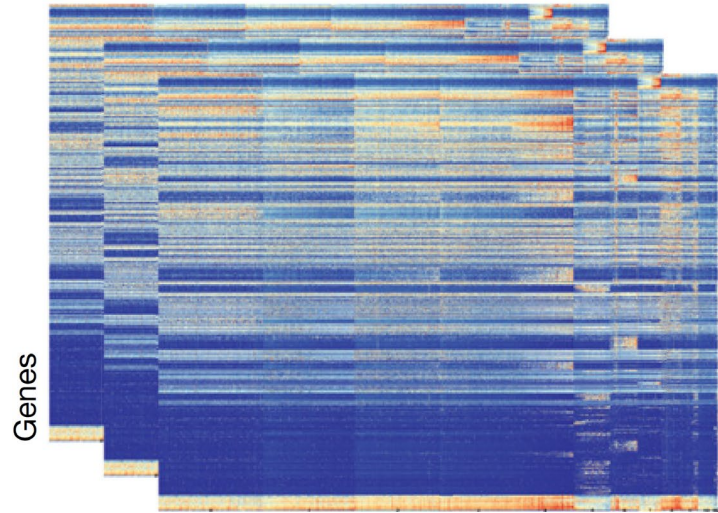
**Poly(dT):** cDNA synthesis

**BC:** Cellular barcode

**UMI:** Unique molecule identifier

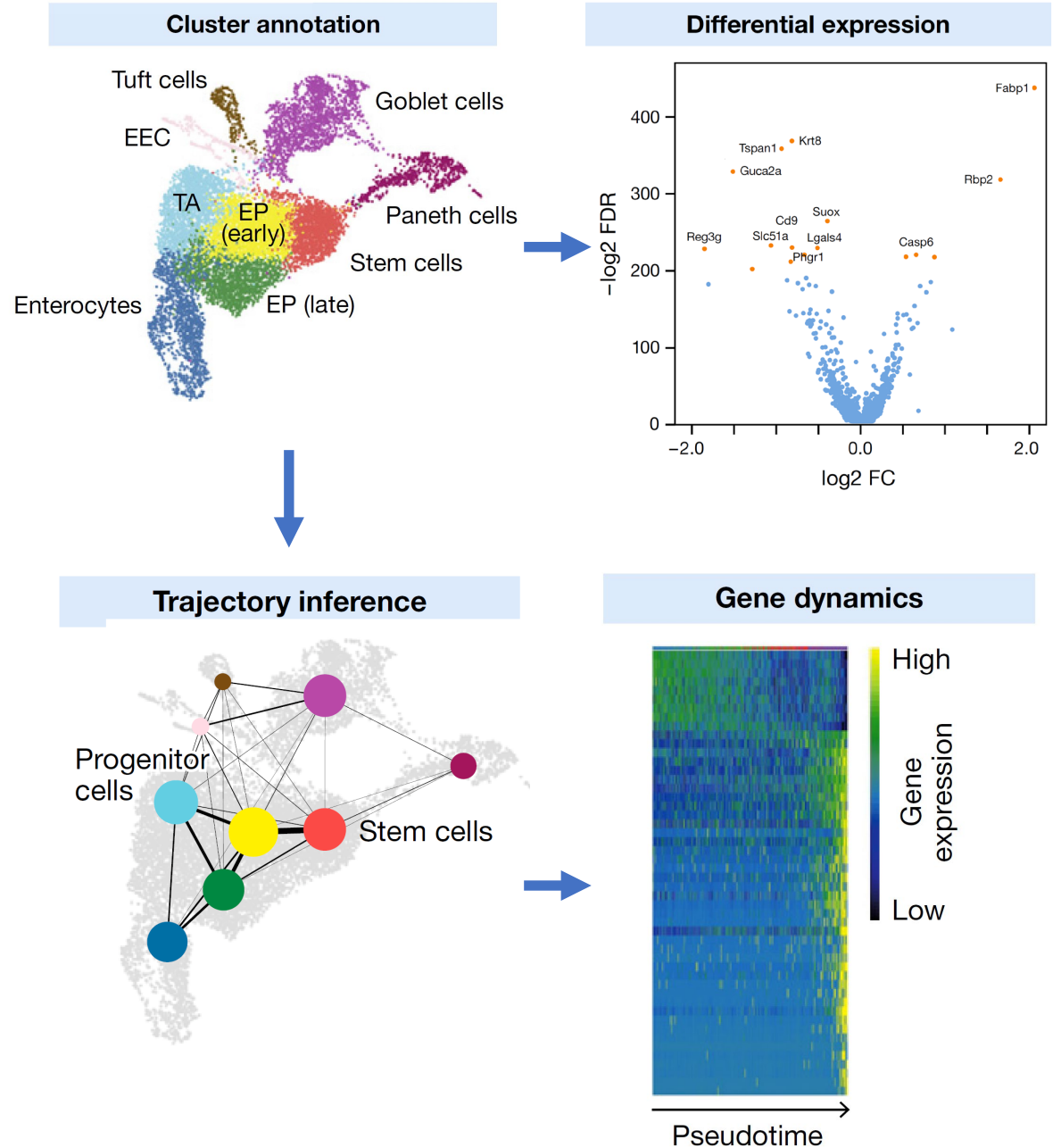
**P5, P7:** Adaptor → Sequencing

# Workflow of scRNA-seq

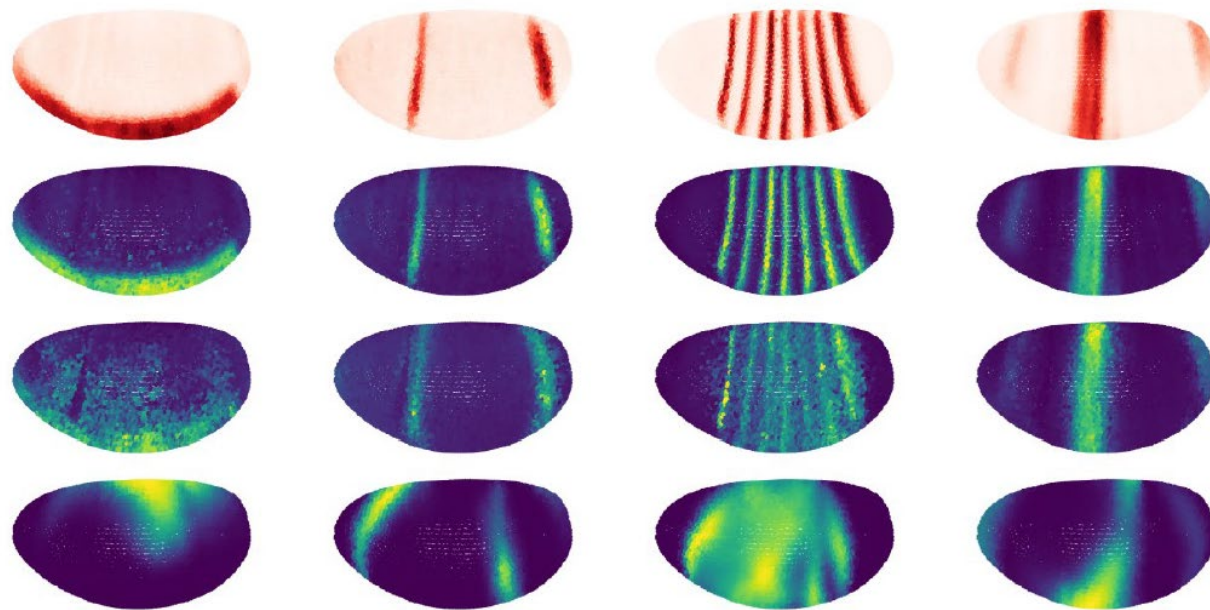
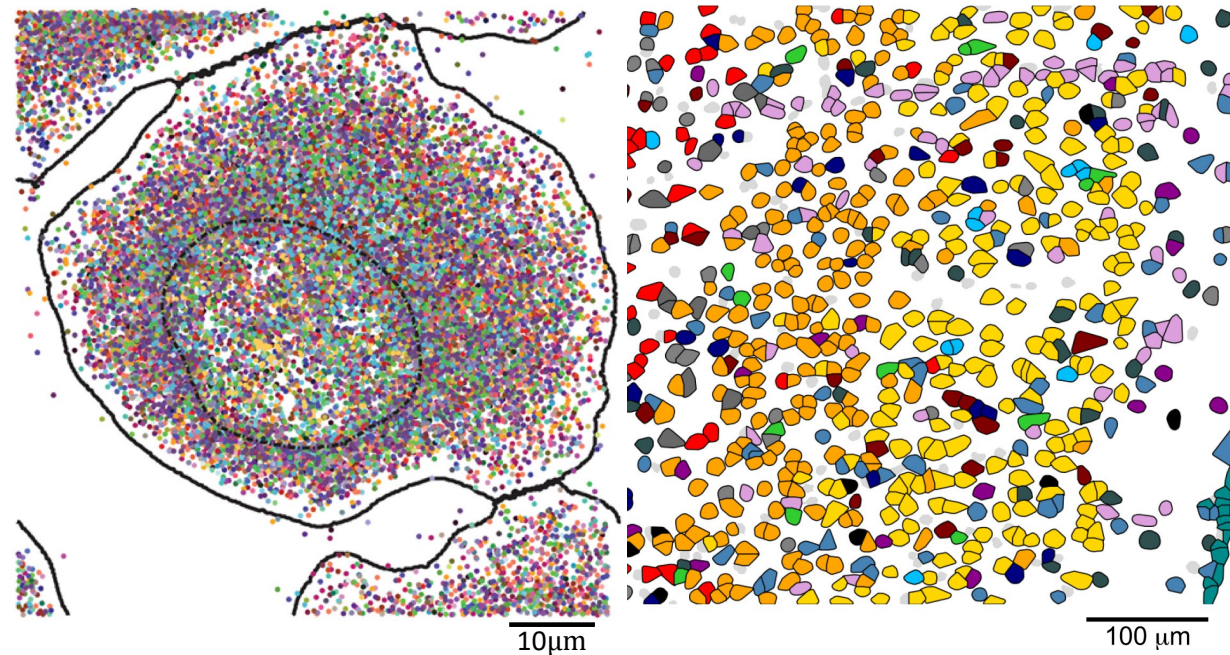
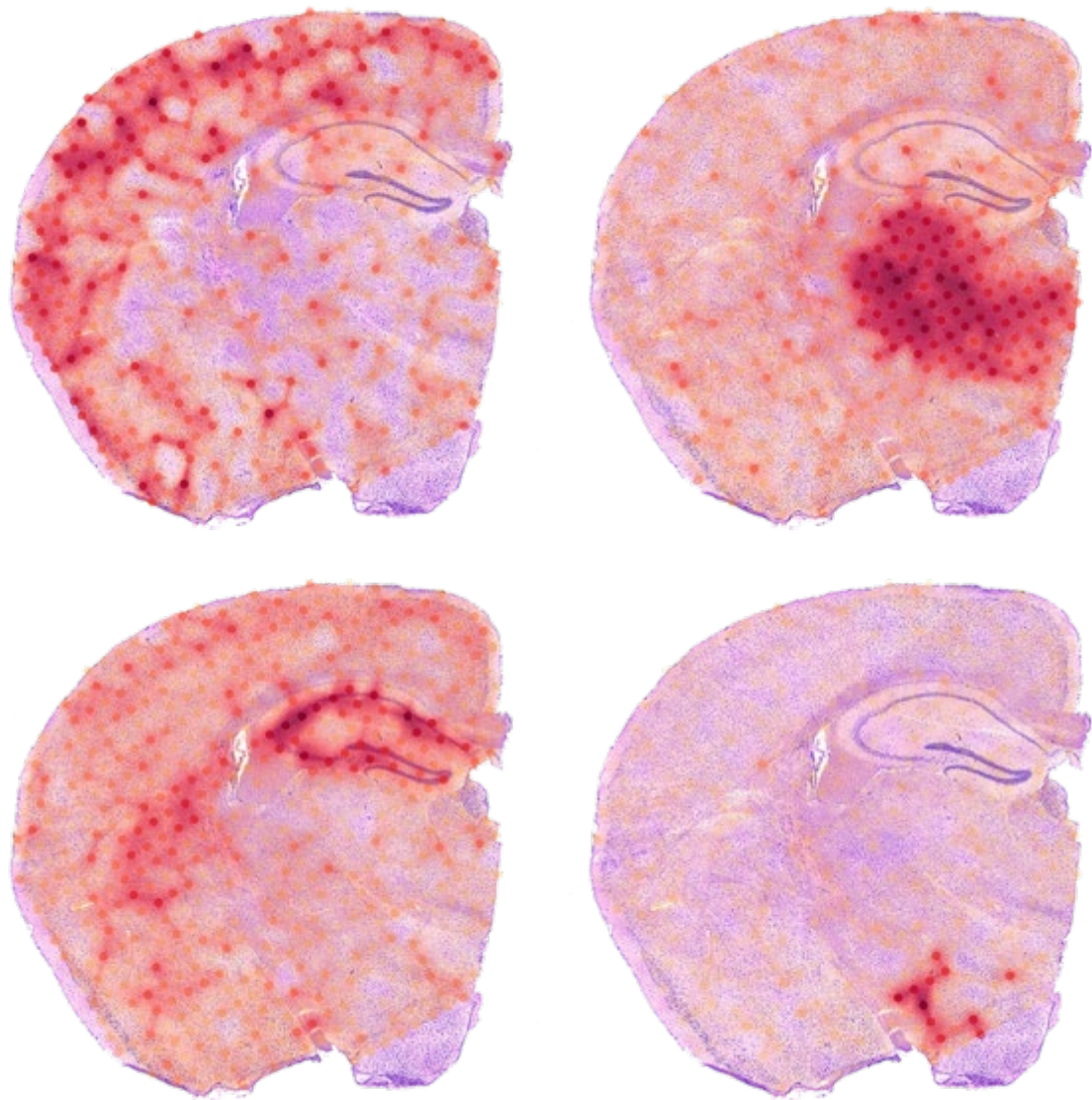


Count matrices

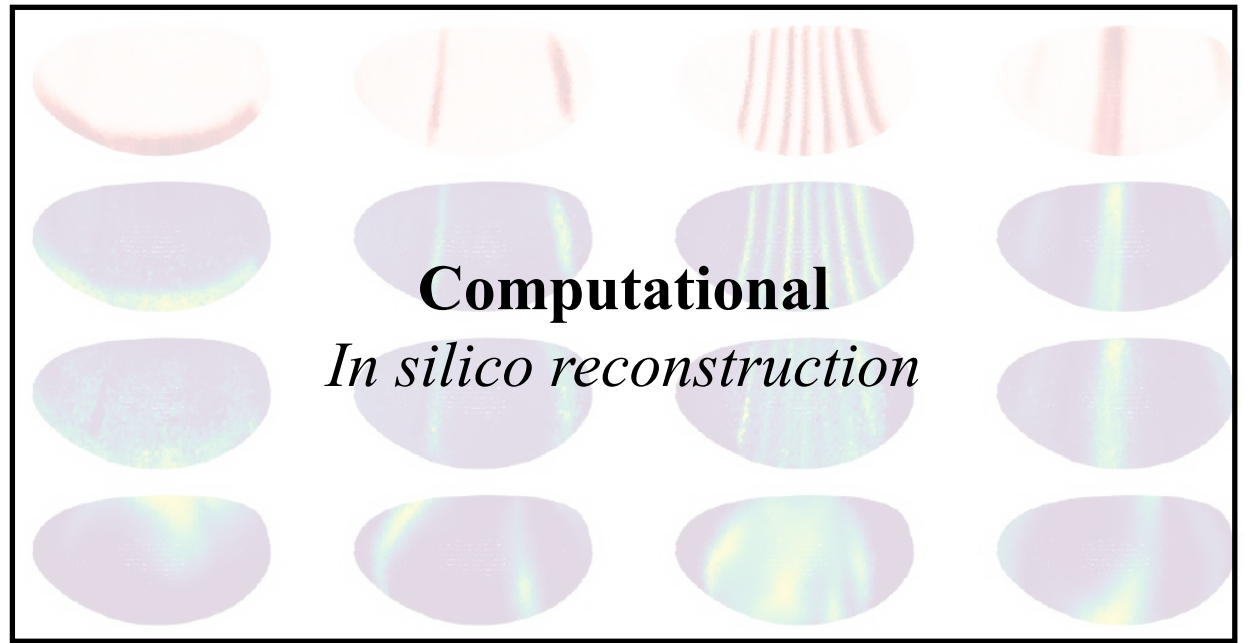
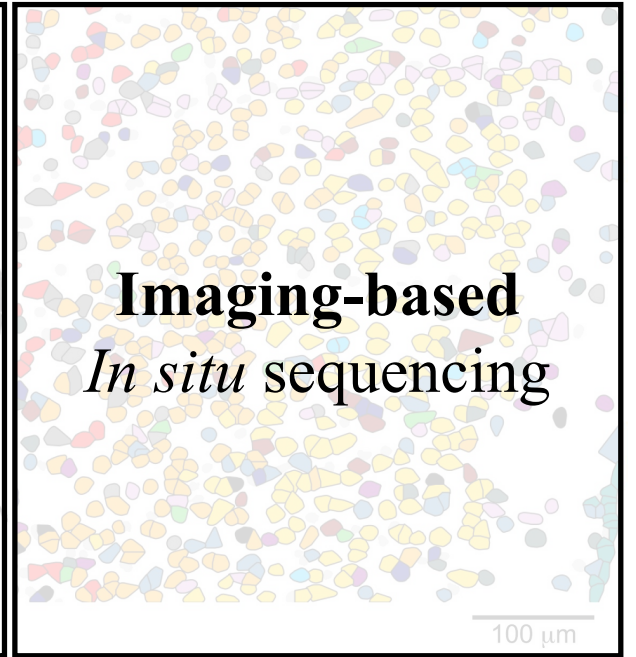
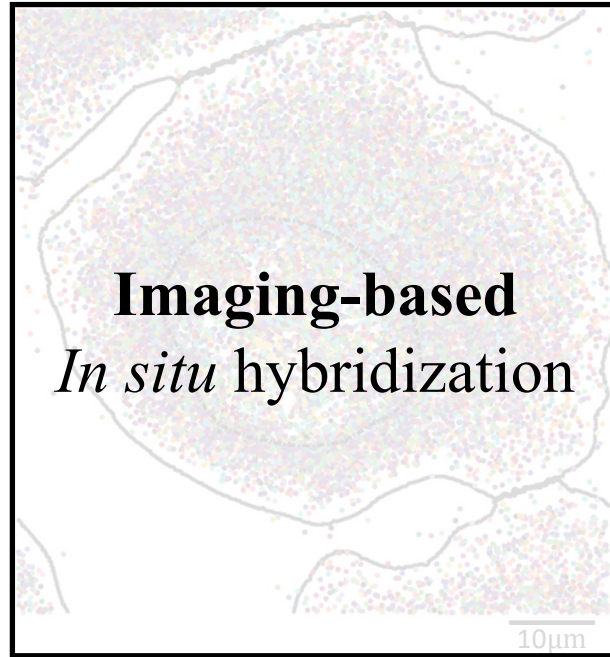
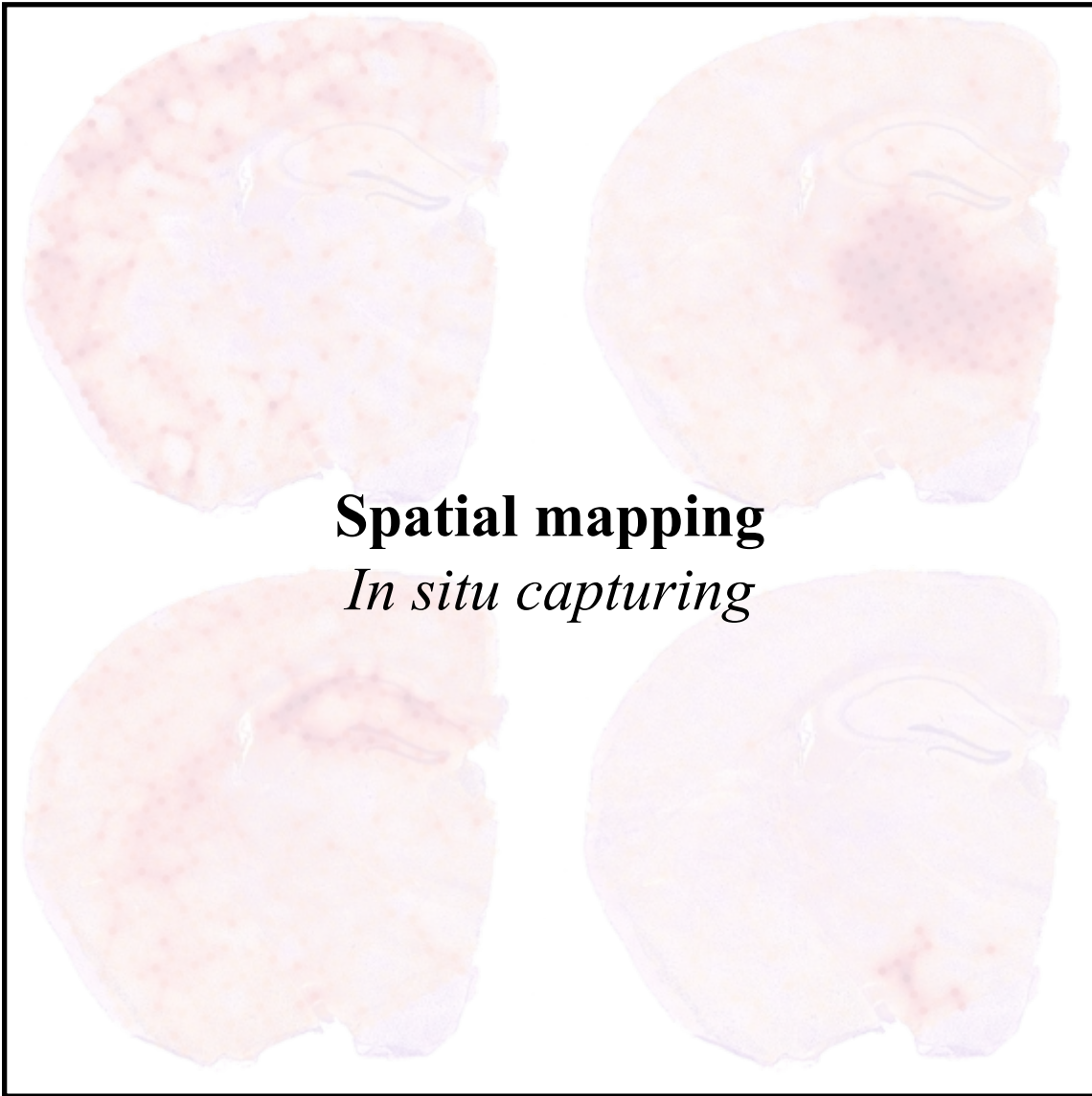
**Spatial information is lost!**



# Spatial transcriptomics

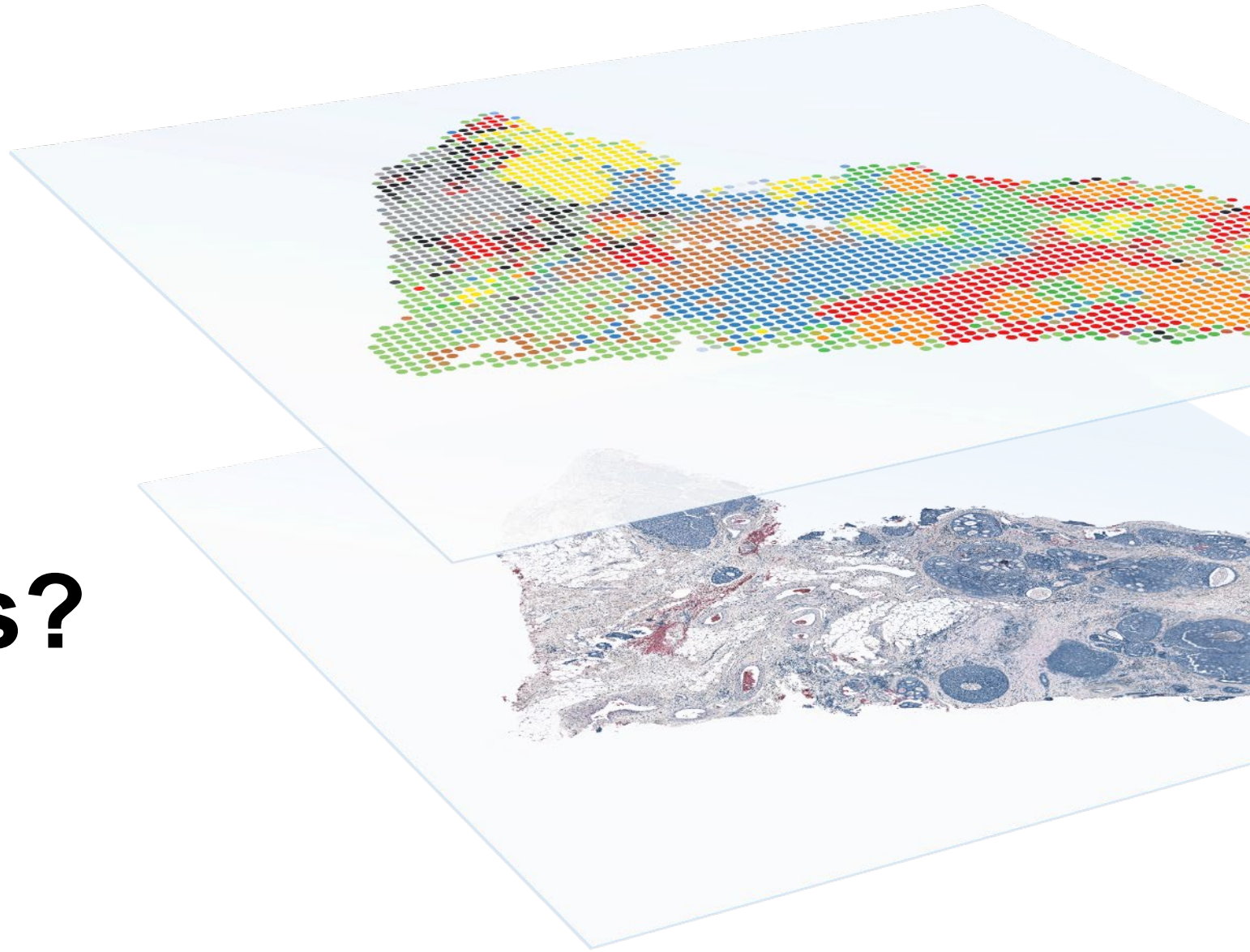


# Spatial transcriptomics



# What is spatial transcriptomics?

Look into four distinctive methods



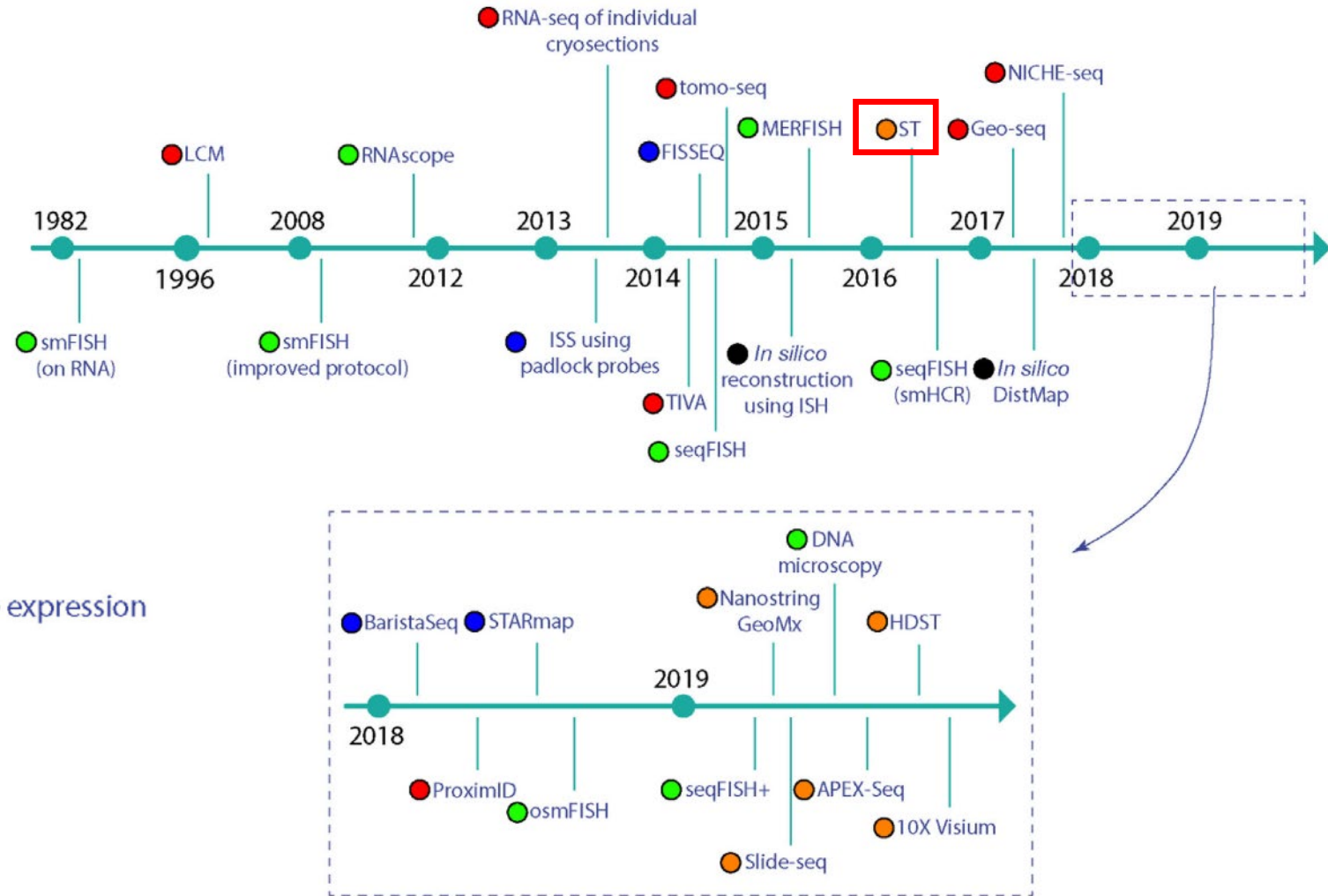
# History timeline

FOCUS | 06 JANUARY 2021

## Method of the Year 2020: spatially resolved transcriptomics

Spatially resolved transcriptomics is our Method of the Year 2020, for its ability to provide valuable insights into the biology of cells and tissues while retaining information about spatial context.

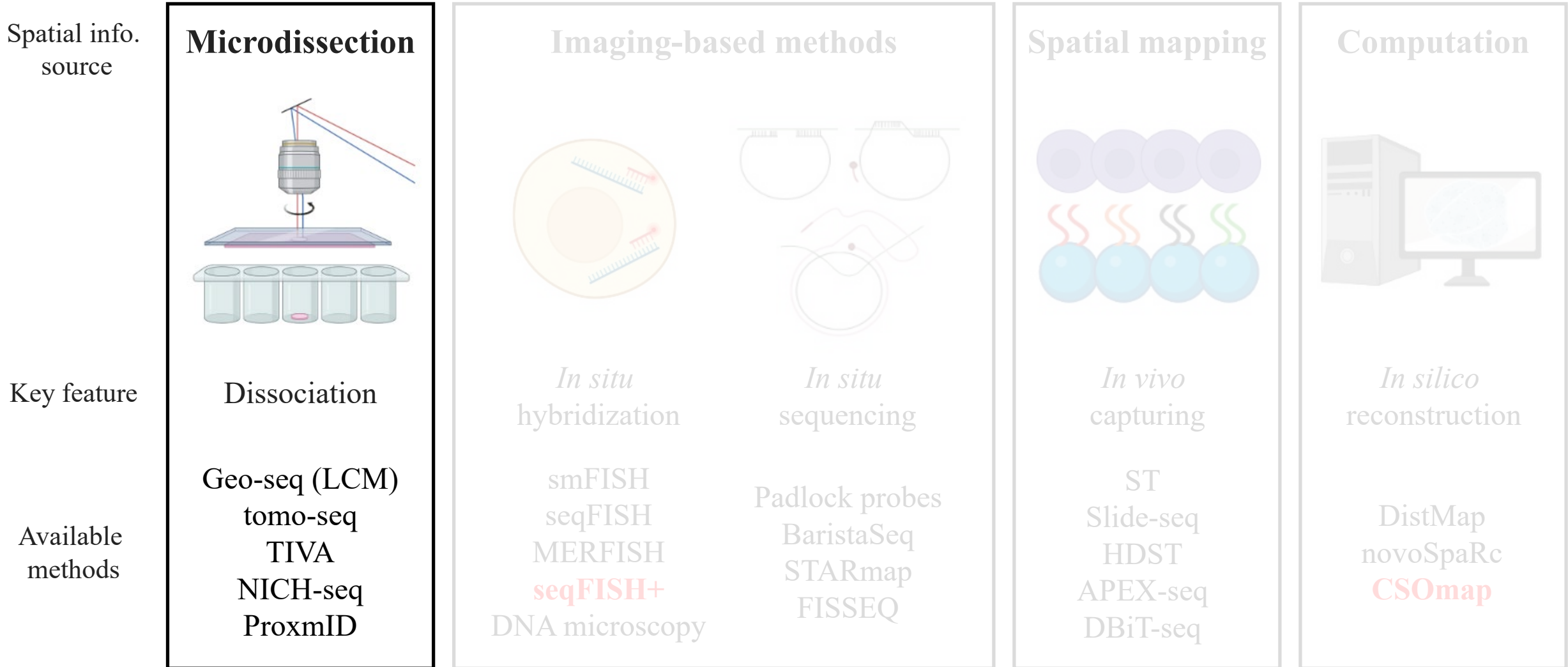
- Section 1. Technologies based on microdissected gene expression
- Section 2. *In situ* hybridization technologies
- Section 3. *In situ* sequencing technologies
- Section 4. *In situ* capturing technologies
- Section 5. *In silico* reconstruction of spatial data



# Four paradigms of spatial transcriptomics

Spatial info. source	Microdissection	Imaging-based methods		Spatial mapping	Computation
Key feature	Dissociation	<i>In situ</i> hybridization	<i>In situ</i> sequencing	<i>In vivo</i> capturing	<i>In silico</i> reconstruction
Available methods	Geo-seq (LCM) tomo-seq TIVA NICH-seq ProxmID	smFISH seqFISH MERFISH <b>seqFISH+</b>	Padlock probes BaristaSeq STARmap FISSEQ	ST Slide-seq HDST DBiT-seq	DistMap novoSpaRc <b>CSOmap</b>

# Four paradigms of spatial transcriptomics

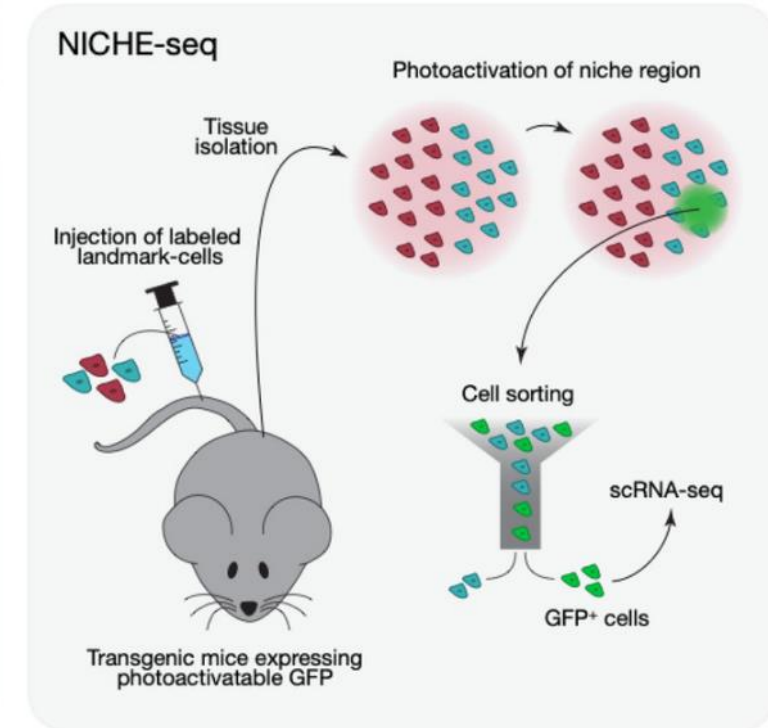
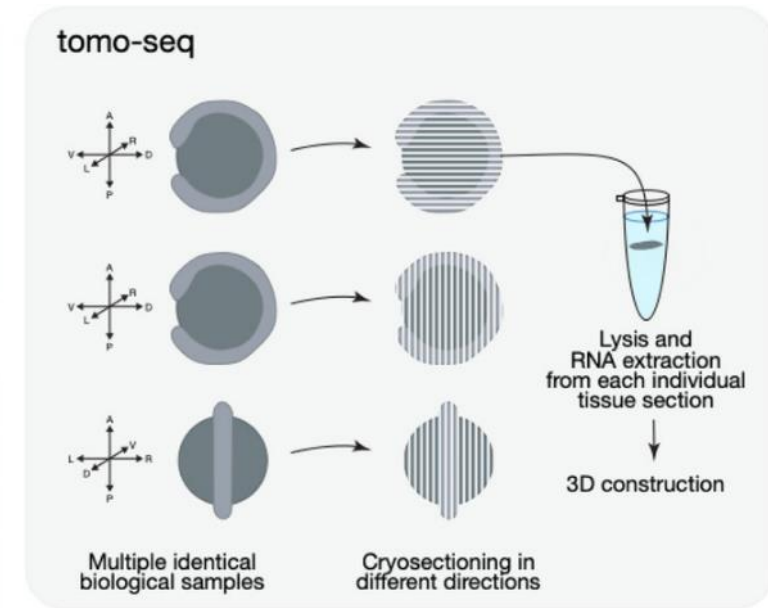
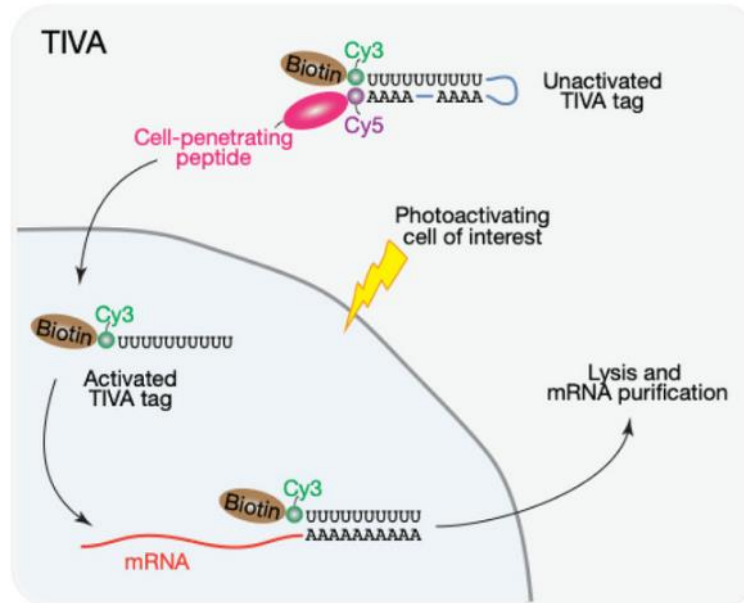
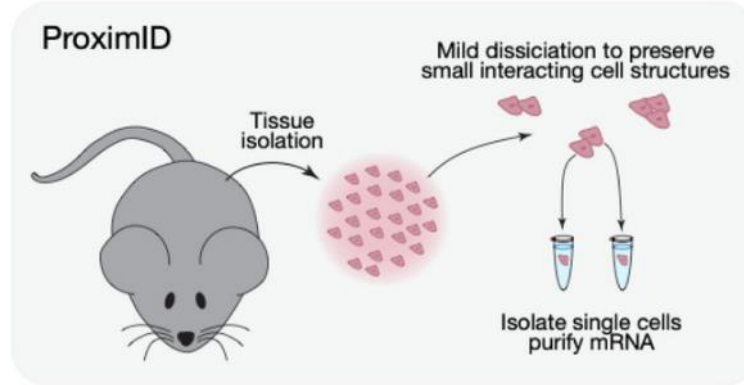
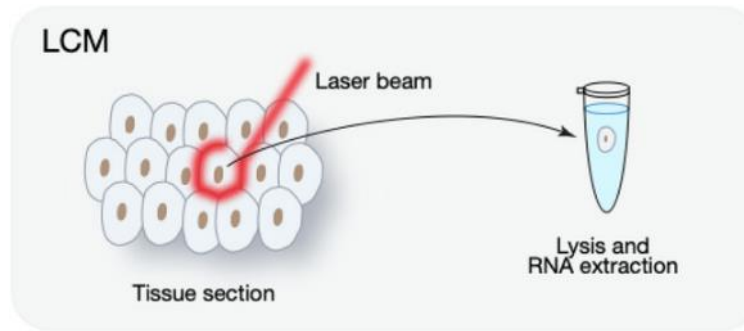




# Microdissection methods isolate individual areas

- + Cellular resolution
- Low cell throughput
- Can't be applied to limited samples

LCM: laser capture microdissection  
 TIVA: transcriptome *in vivo* analysis

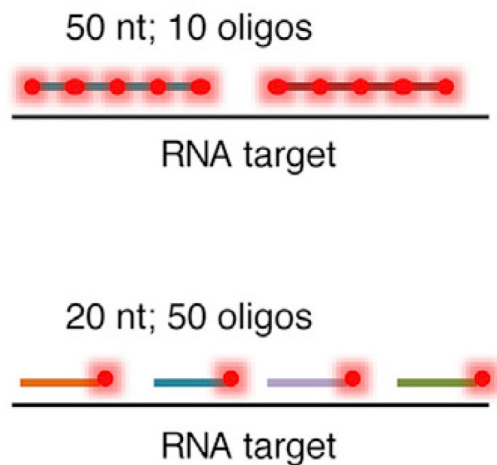


# Four paradigms of spatial transcriptomics

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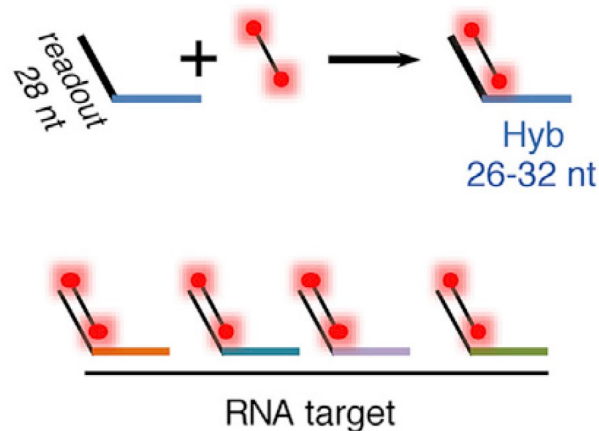
# DNA probes hybridization localizes cellular transcripts

Probes with  
fluorophores



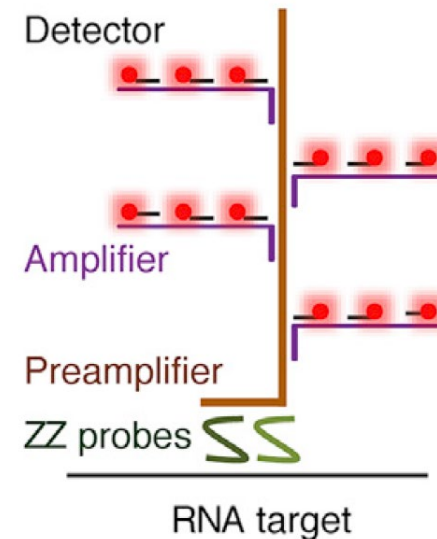
↓  
smFISH

Probes with  
secondary probes



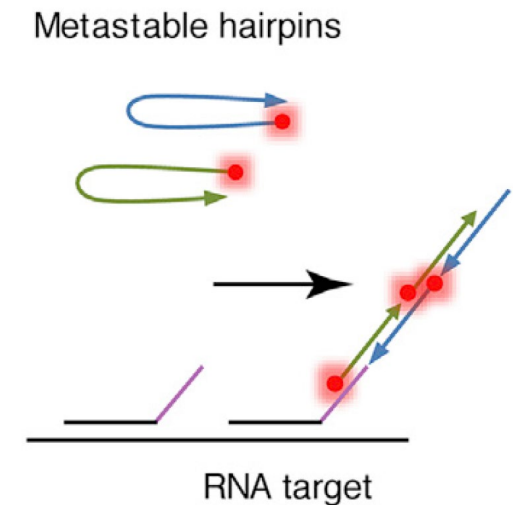
↓  
smiFISH

Probes with  
branched DNA



↓  
RNAscope

Probes with  
chain reaction



↓  
smHCR

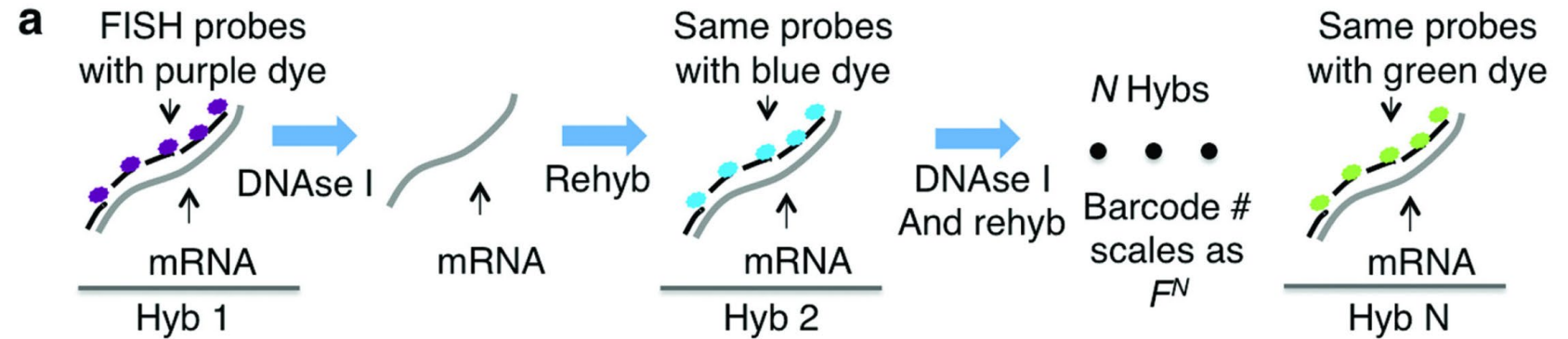
**sm**: single molecule  
**smi**: single molecule inexpensive  
**HCR**: hybridization chain reaction

+ Subcellular resolution – Low transcript throughput

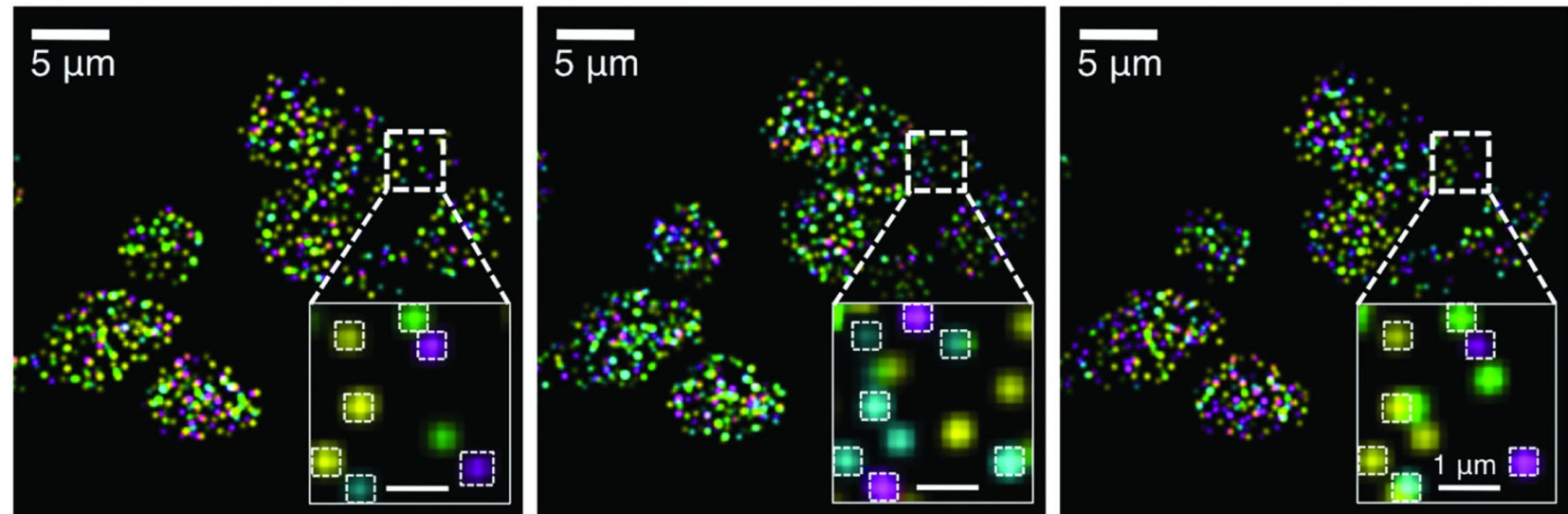
# seqFISH realized multiplexing by multiple hybridization rounds



Prof. Long Cai  
Caltech, USA

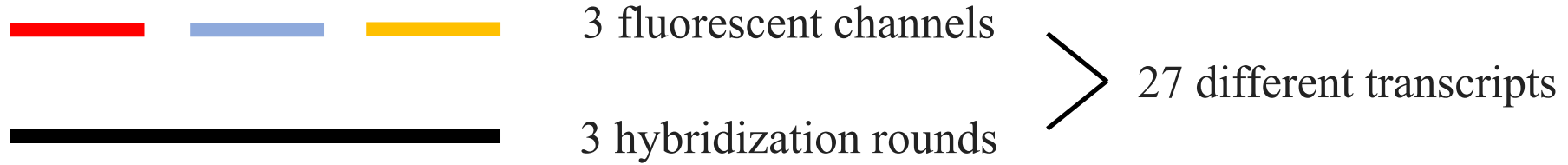


**b** Composite four-color FISH images  
Hybridization 1 – probe set 1    Hybridization 2 – probe set 2    Hybridization 3 – probe set 1

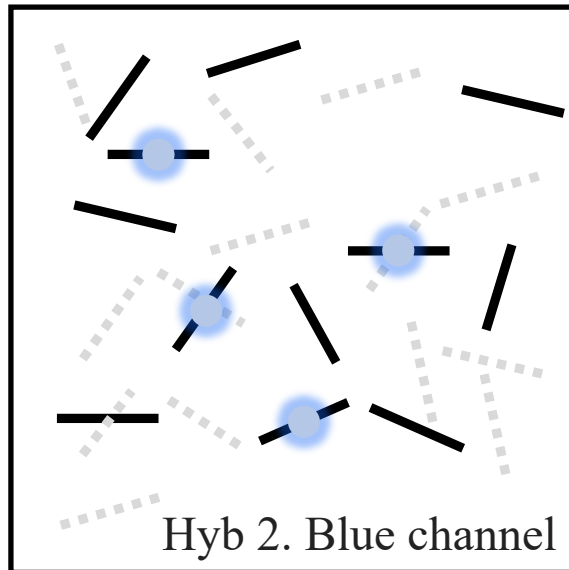


- + Subcellular resolution
- + Highly multiplexed
- Optical crowding

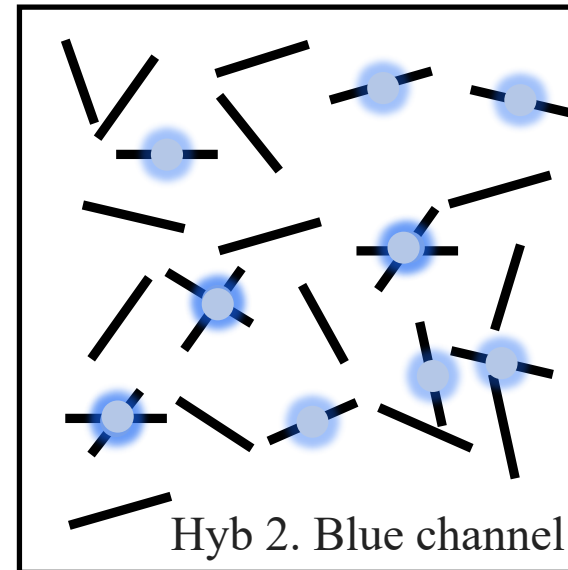
# Optical crowding problem presents a challenge to multiplexing



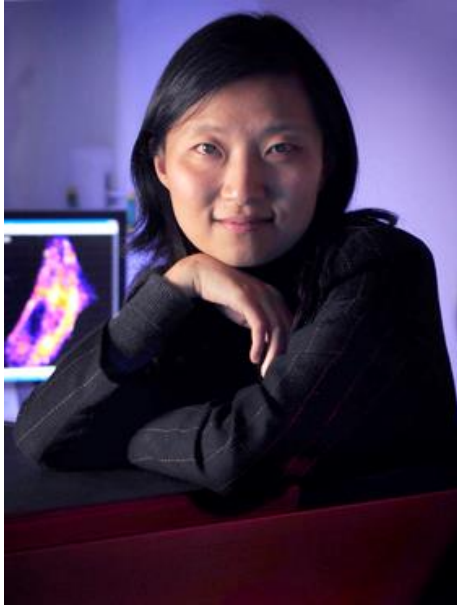
12 targets (no crowding)



27 targets (crowding)



# MERFISH and seqFISH+ resolved optical crowding by an encoding-readout labeling strategy



Prof. Xiaowei Zhuang  
Harvard University

## RESEARCH ARTICLE SUMMARY

### RNA IMAGING

## Spatially resolved, highly multiplexed RNA profiling in single cells

Kok Hao Chen,<sup>1\*</sup> Alistair N. Boettiger,<sup>1\*</sup> Jeffrey R. Moffitt,<sup>1\*</sup>  
Siyuan Wang,<sup>1</sup> Xiaowei Zhuang<sup>1,2+</sup>

## LETTER

<https://doi.org/10.1038/s41586-019-1049-y>

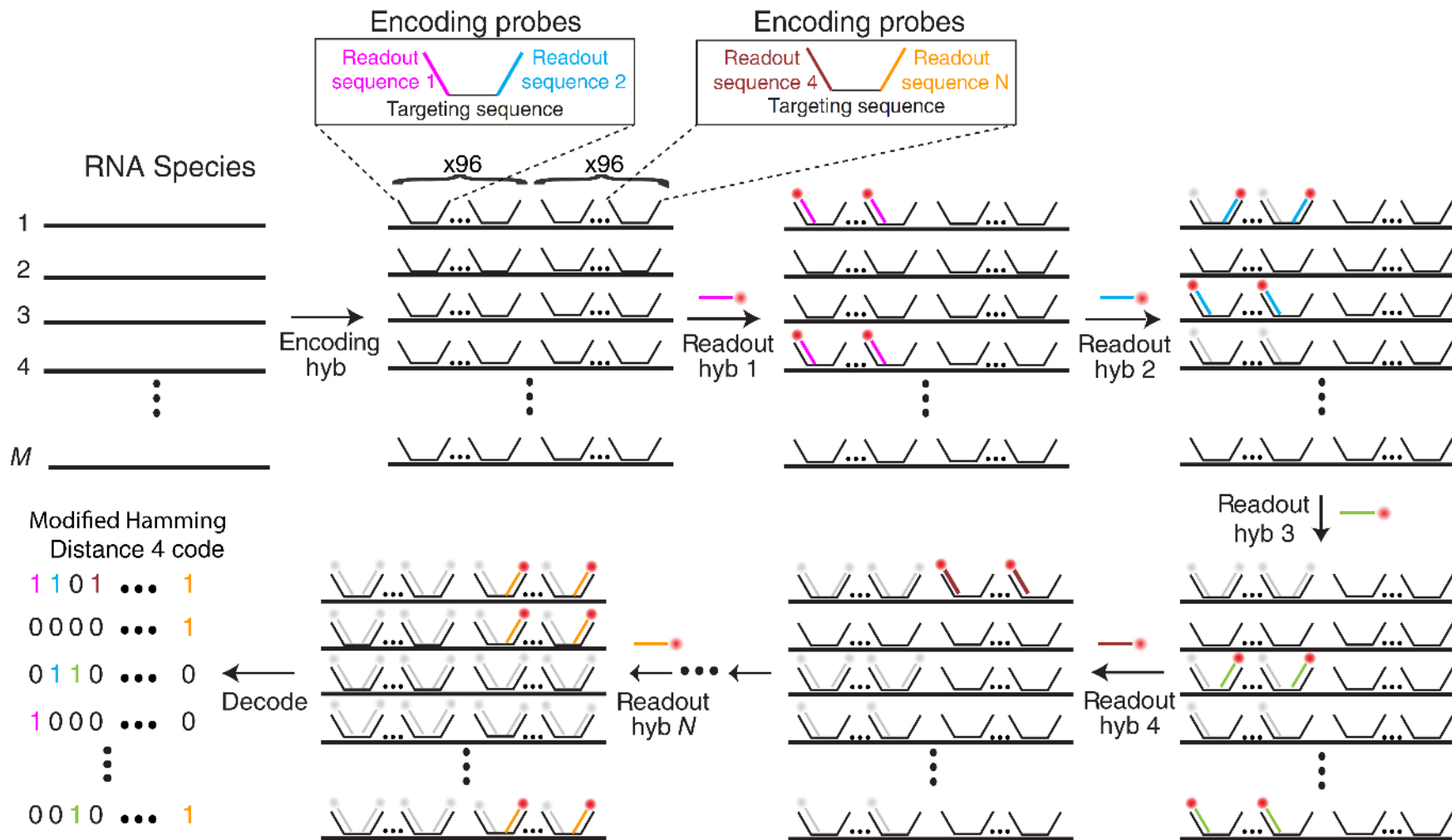
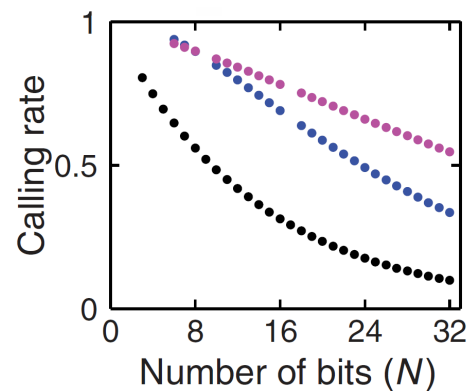
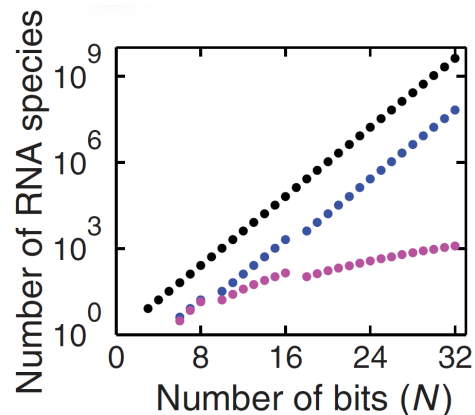
## Transcriptome-scale super-resolved imaging in tissues by RNA seqFISH+

Chee-Huat Linus Eng<sup>1</sup>, Michael Lawson<sup>2</sup>, Qian Zhu<sup>3</sup>, Ruben Dries<sup>3</sup>, Noushin Koulana<sup>2</sup>, Yodai Takei<sup>2</sup>, Jina Yun<sup>2</sup>, Christopher Cronin<sup>2</sup>, Christoph Karp<sup>2</sup>, Guo-Cheng Yuan<sup>3</sup> & Long Cai<sup>2\*</sup>

Prof. Long Cai  
Caltech



# Multiplexed error-robust FISH



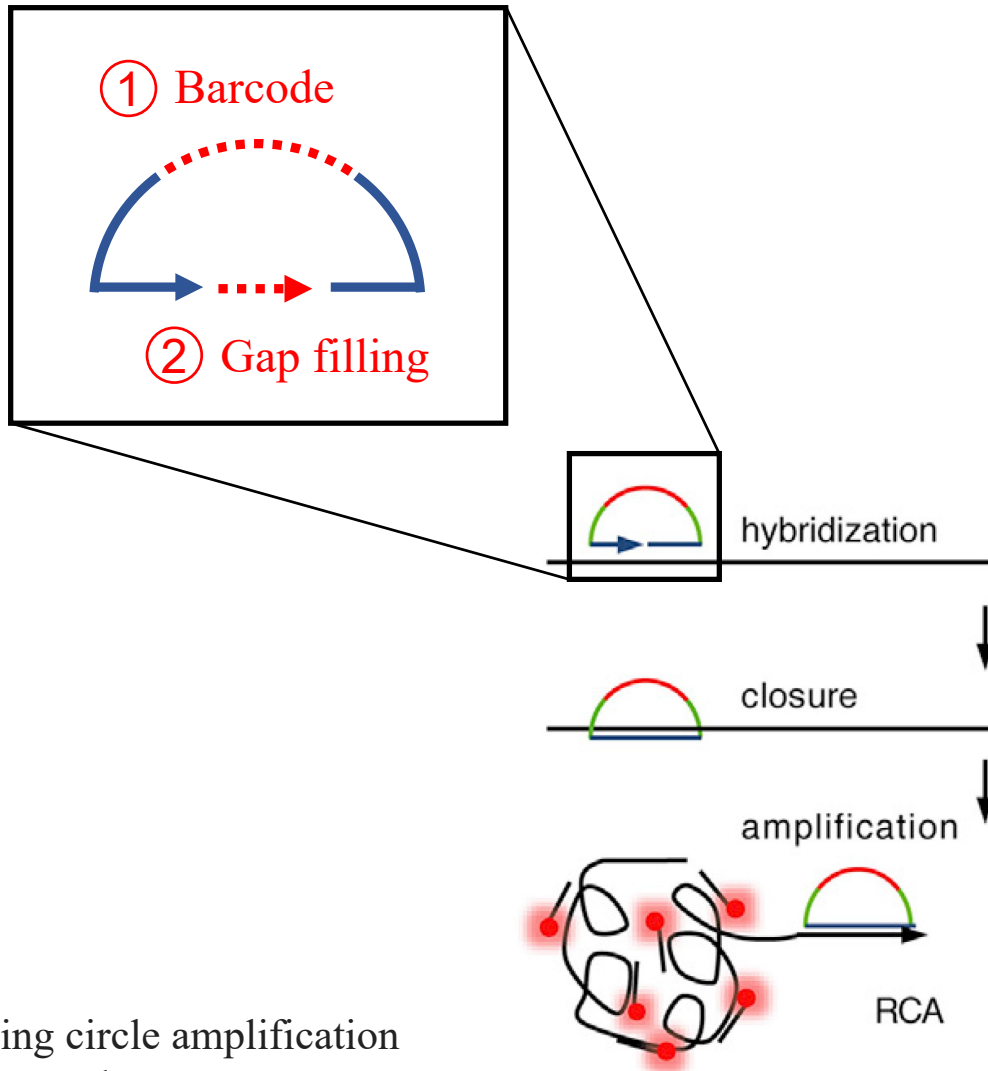
Modified HD4 code

# Four paradigms of spatial transcriptomics

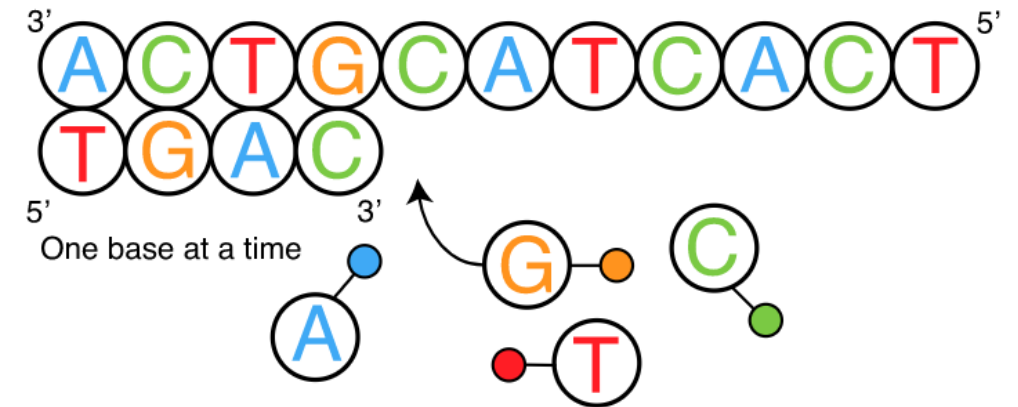
Spatial info. source	Microdissection	Imaging-based methods		Spatial mapping	Computation
Key feature	Dissociation	<i>In situ</i> hybridization	<i>In situ</i> sequencing	<i>In vivo</i> capturing	<i>In silico</i> reconstruction
Available methods	Geo-seq (LCM) tomo-seq TIVA NICH-seq ProxmID	smFISH seqFISH MERFISH <b>seqFISH+</b>	Padlock probes BaristaSeq STARmap FISSEQ	ST Slide-seq HDST DBiT-seq	DistMap novoSpaRc <b>CSOmap</b>



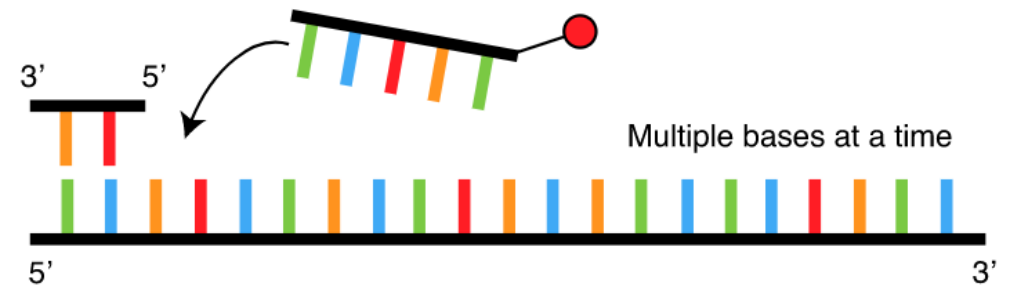
# Padlock probes hybridization localizes cellular transcripts



## Sequencing by Synthesis



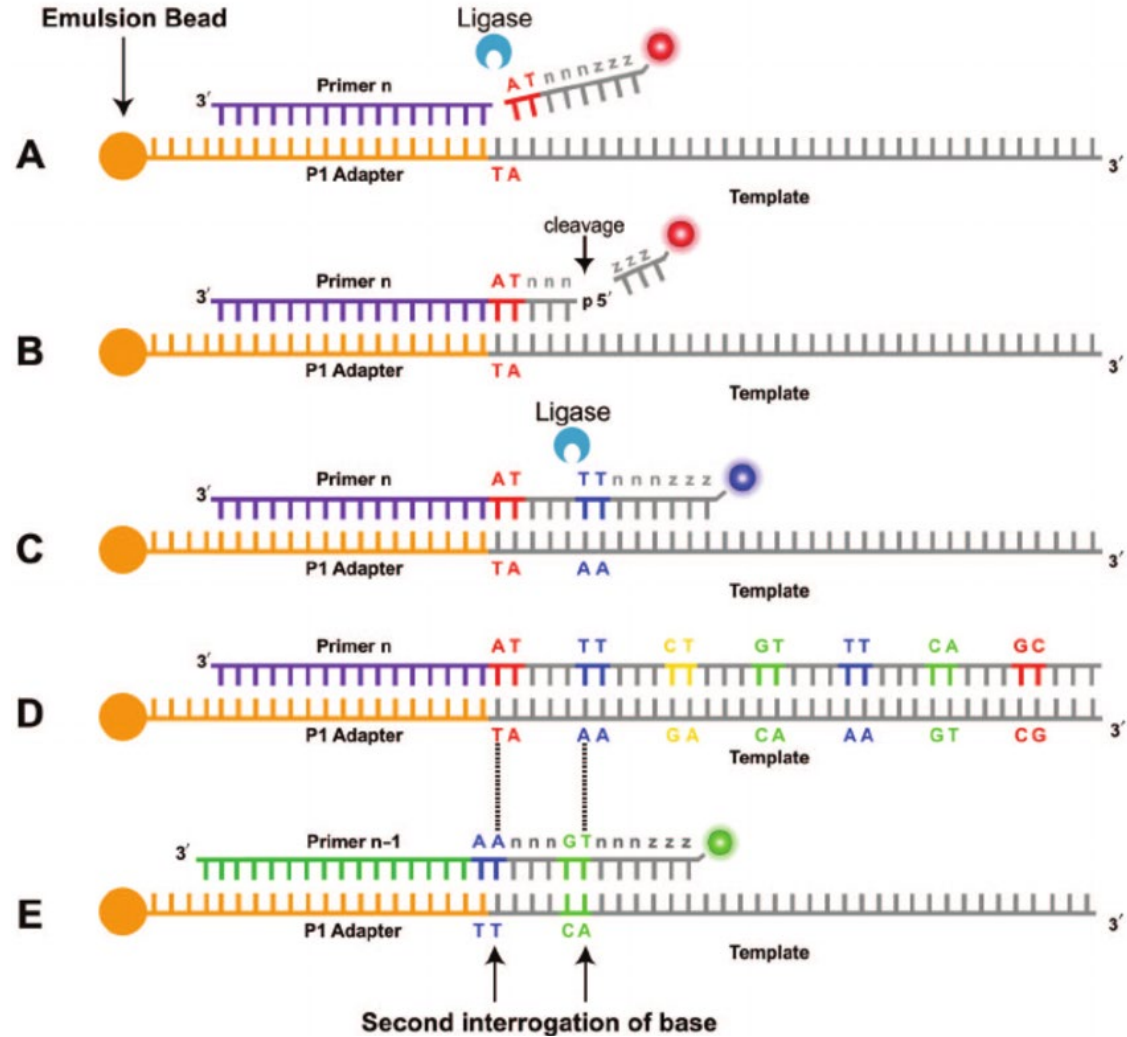
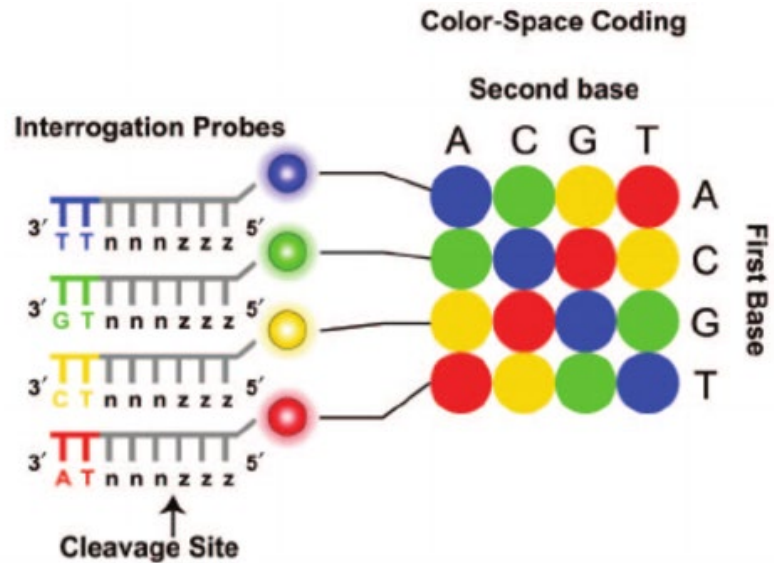
## Sequencing by Ligation



RCA: rolling circle amplification  
RCP: RCA product

# Sequencing by oligonucleotide ligation and detection

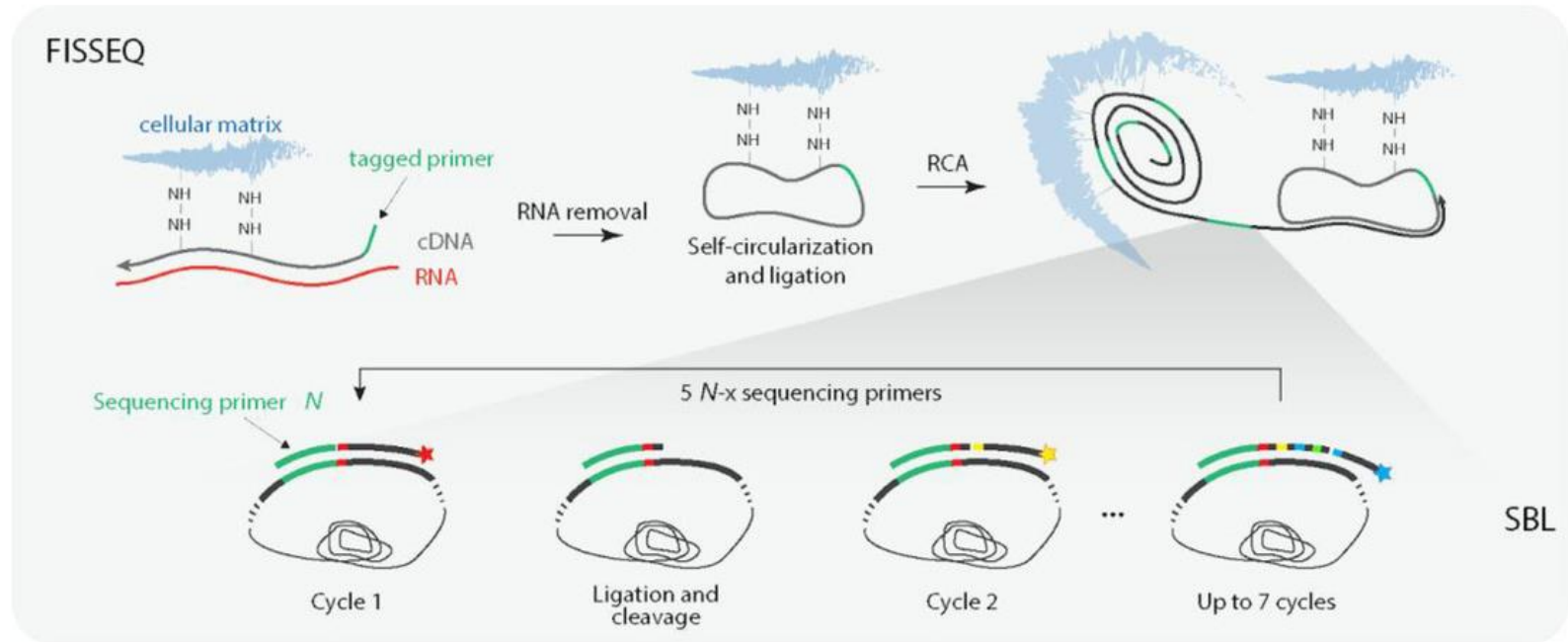
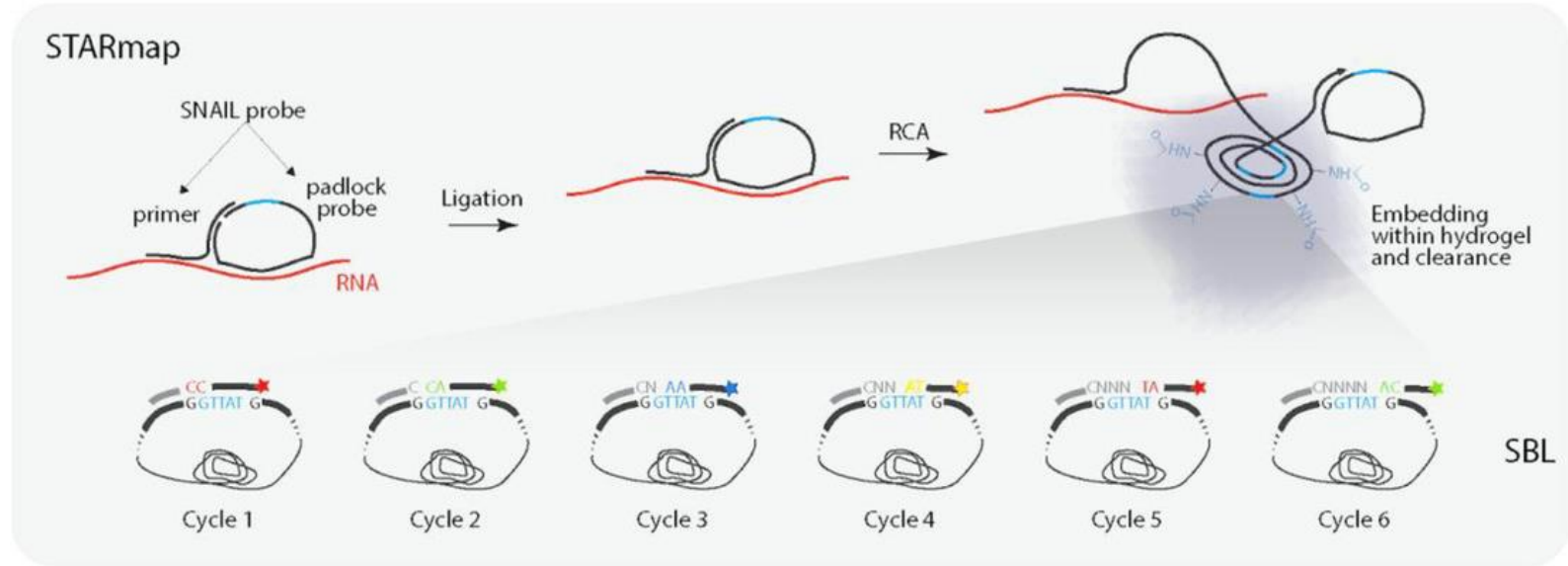
SOLiD sequencing



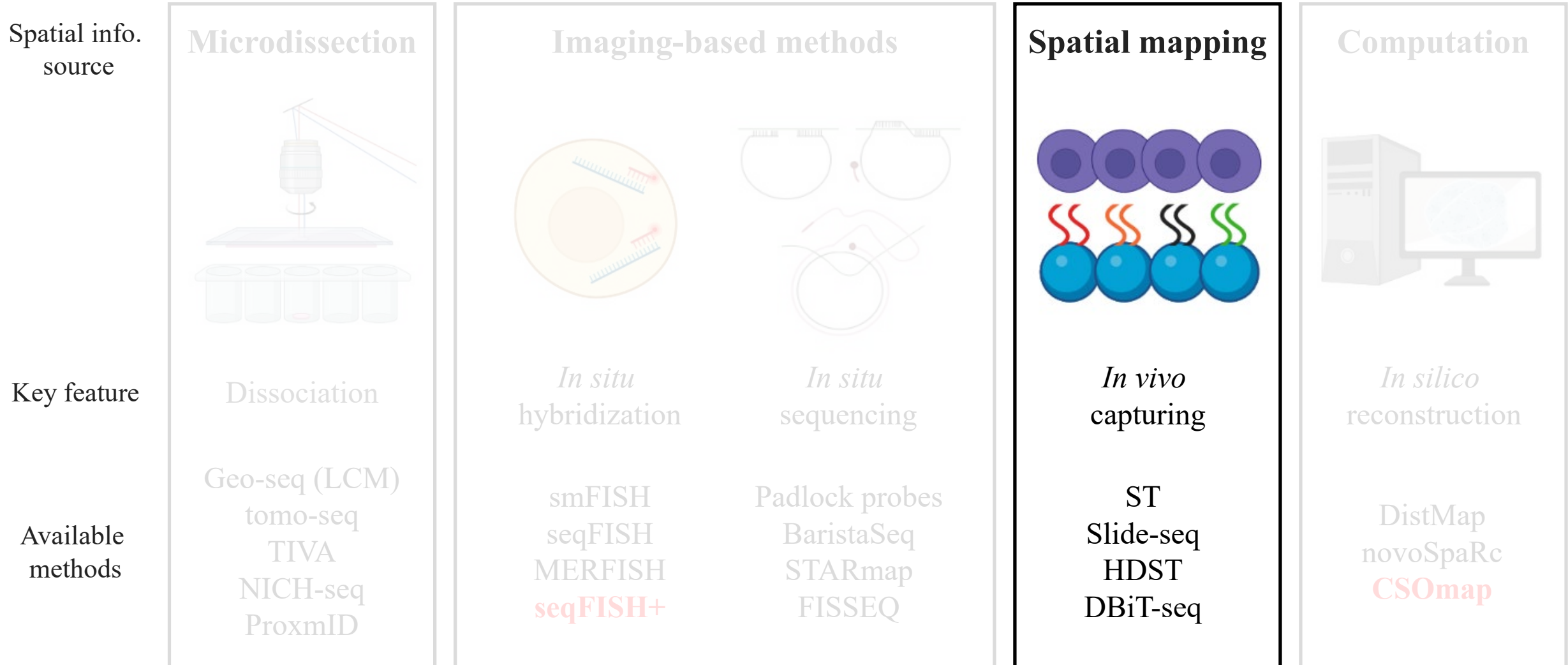
# Targeted or untargeted *in situ* sequencing

- + Subcellular resolution
- Limit quantity
- Some only applied in cultured cell

**FISSEQ**: fluorescent *in situ* RNA sequencing  
**STARmap**: spatially-resolved transcript amplicon readout mapping



# Four paradigms of spatial transcriptomics



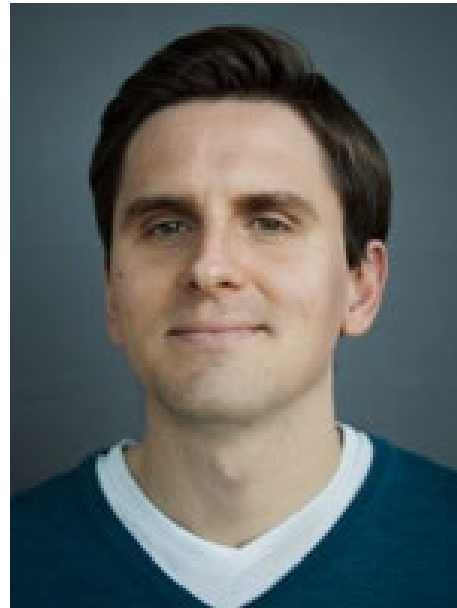
# Pioneers of the field

Spatial transcriptomics (ST)  
Slide-seq

High-definition spatial  
transcriptomics (HDST)



Prof. Joakim Lundeberg  
KTH, Sweden



Prof. Patrik Ståhl  
KTH, Sweden



Prof. Evan Macosko  
Broad Institute, USA

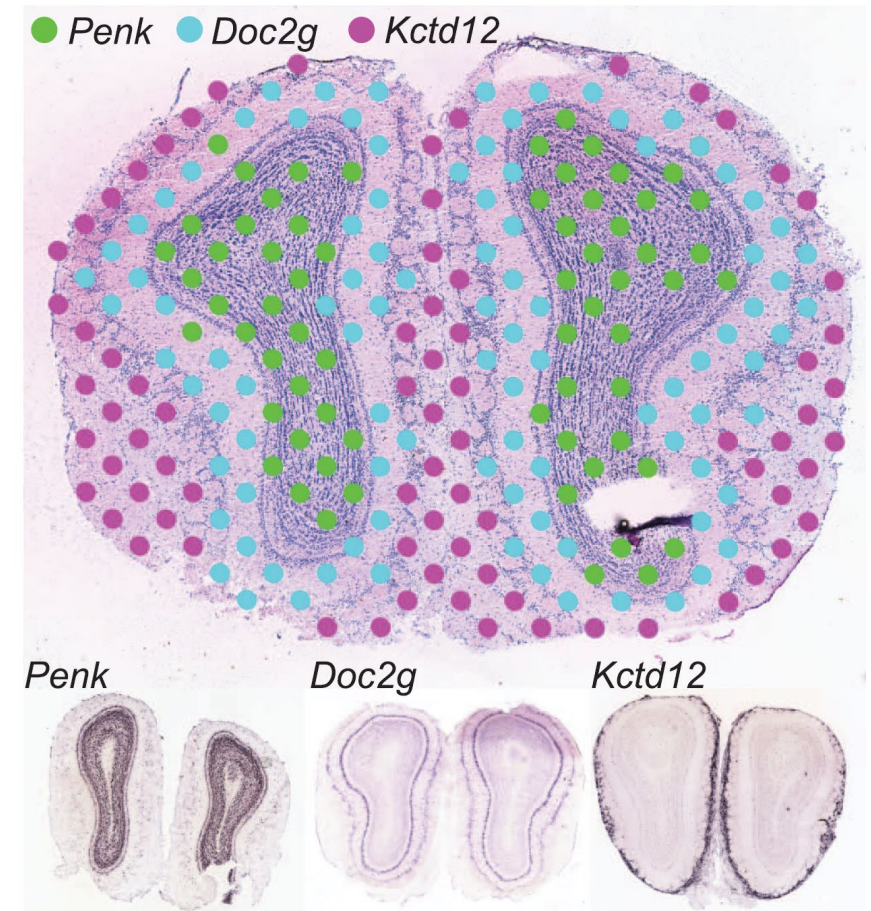
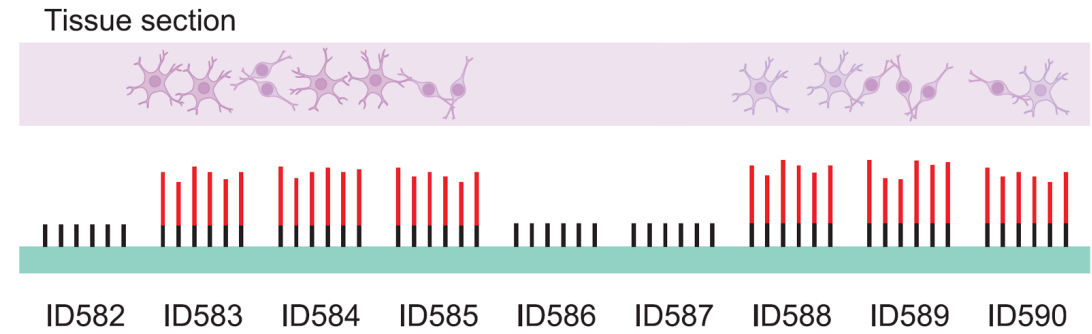
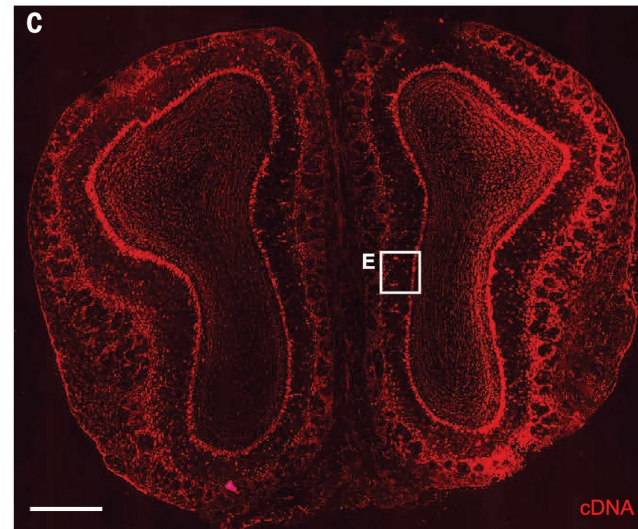
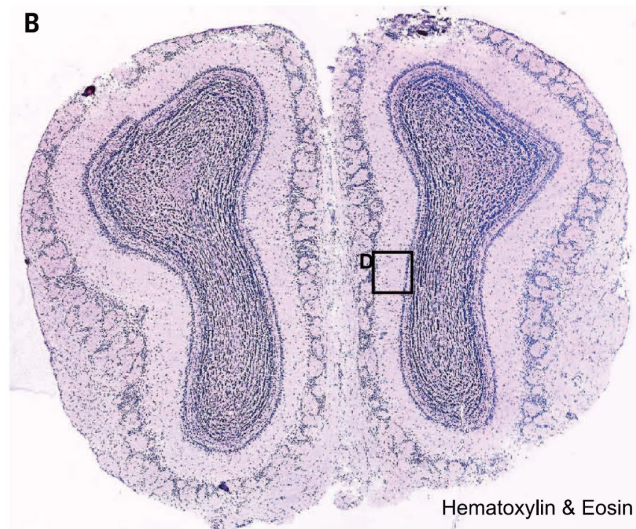
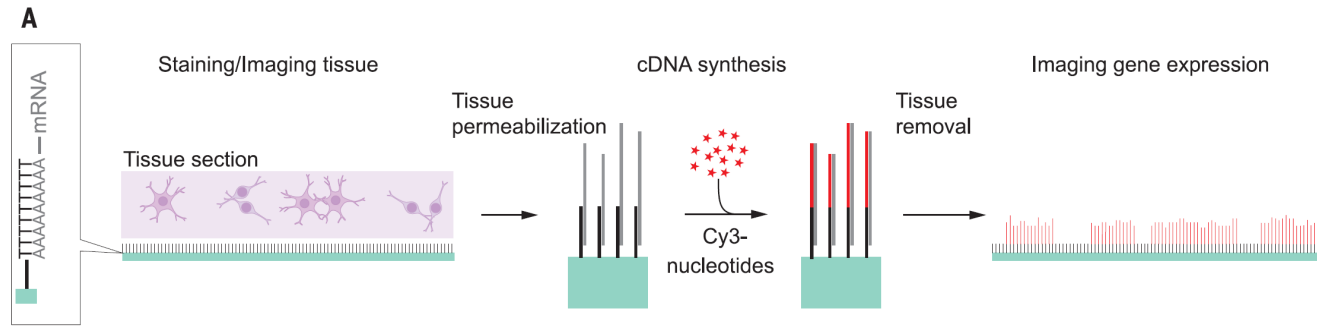


Prof. Fei Chen  
Broad Institute, USA

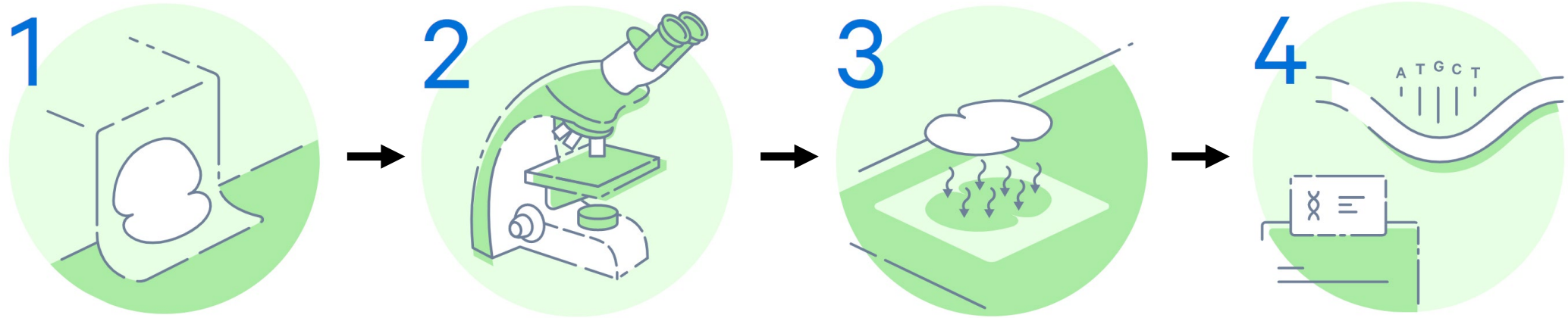
## Spatial mapping methods

# Spatial transcriptomics (ST)

+ Whole mRNA – 100  $\mu$ m resolution



# Workflow of spatial mapping methods



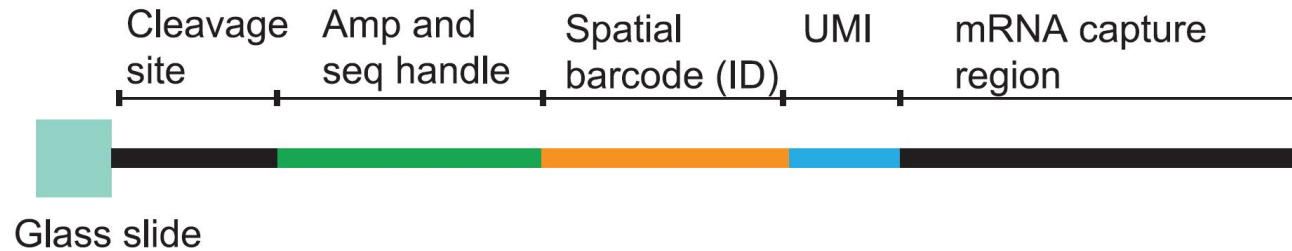
Sample preparation

Stain and imaging

Permeabilize and Library construction

Sequencing

Surface probe



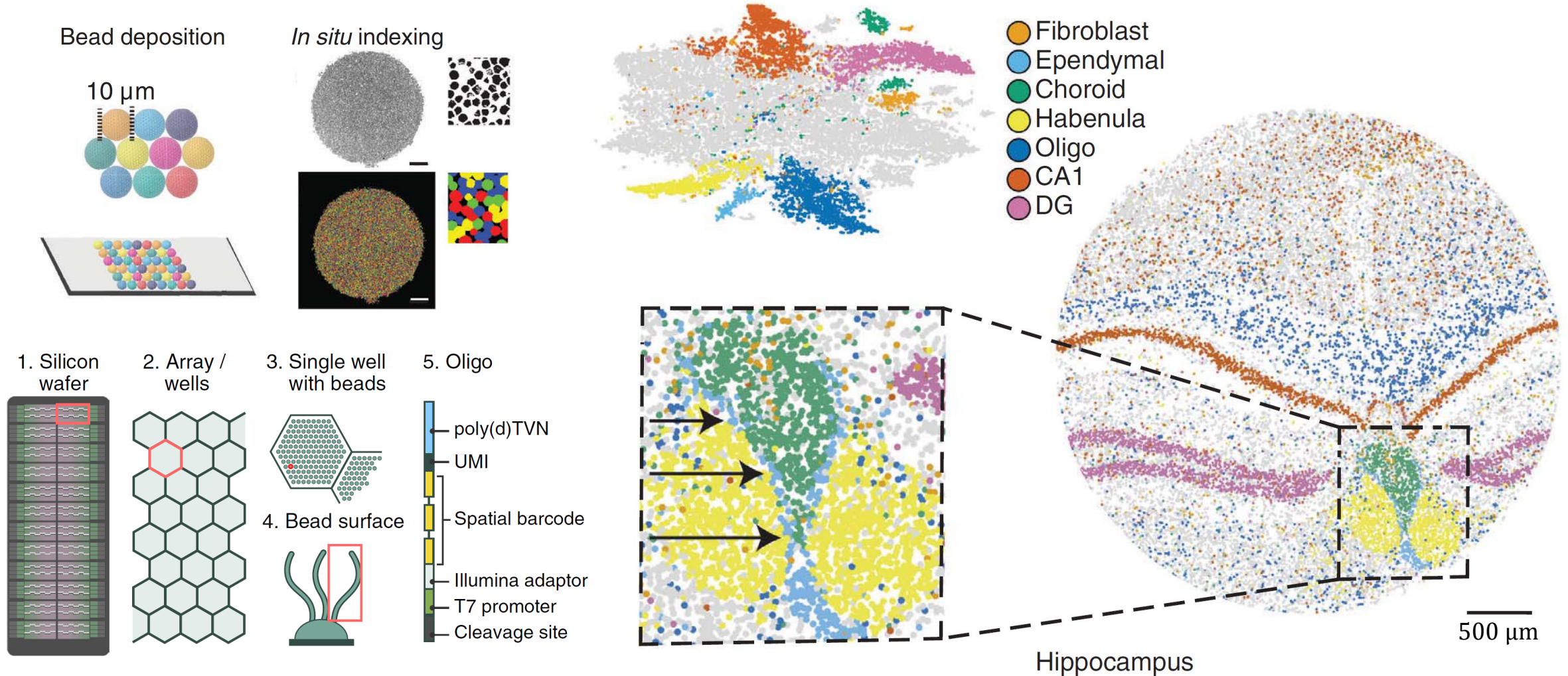
**Spatial barcode:** spatial ID

**UMI:** Unique molecule identifier

**AmpSeq:** PCR + Sequencing

# Slide-seq and High-definition spatial transcriptomics (HDST)

+ Cellular resolution – Low efficiency – Require scRNA-seq data

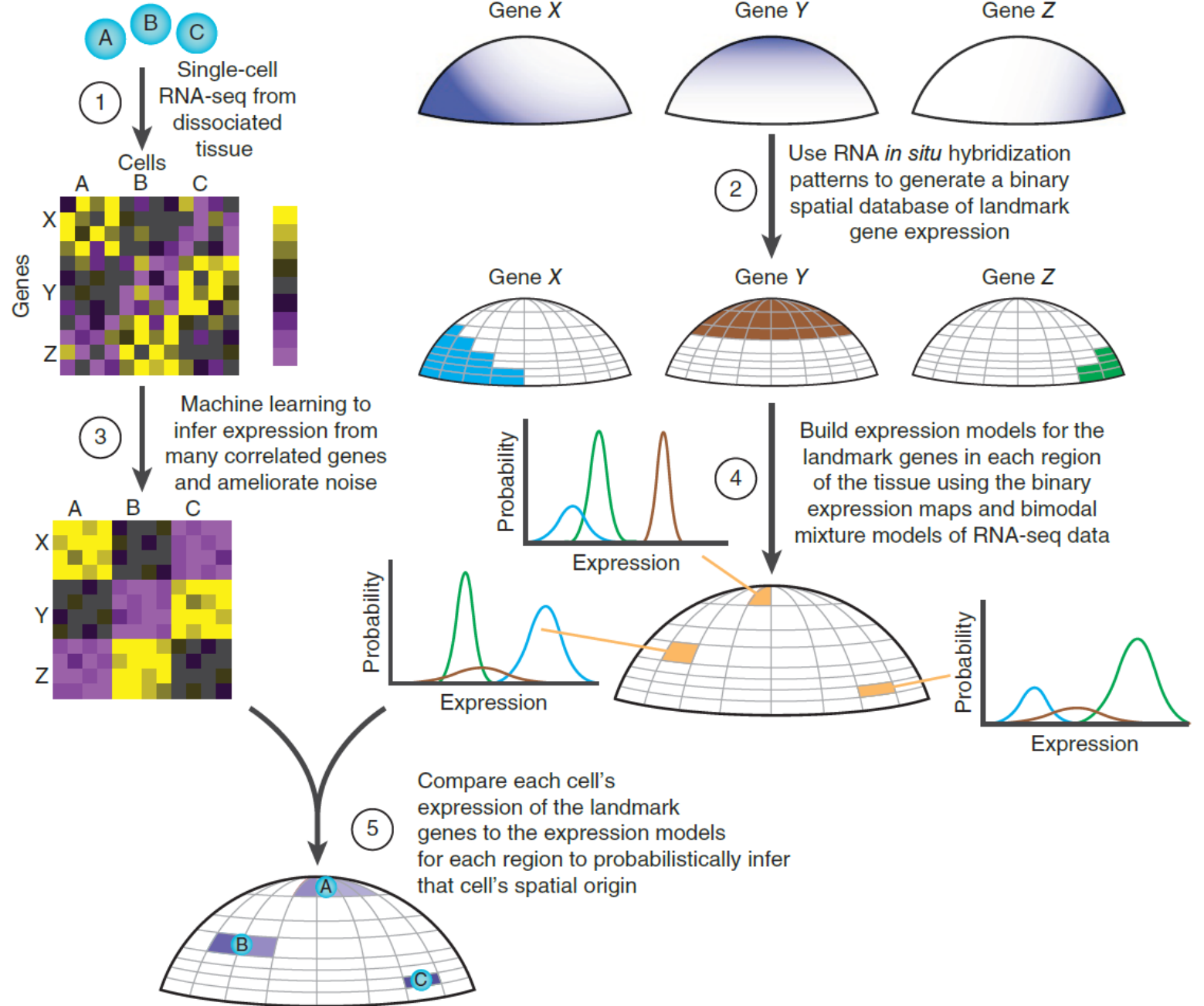




# Four paradigms of spatial transcriptomics

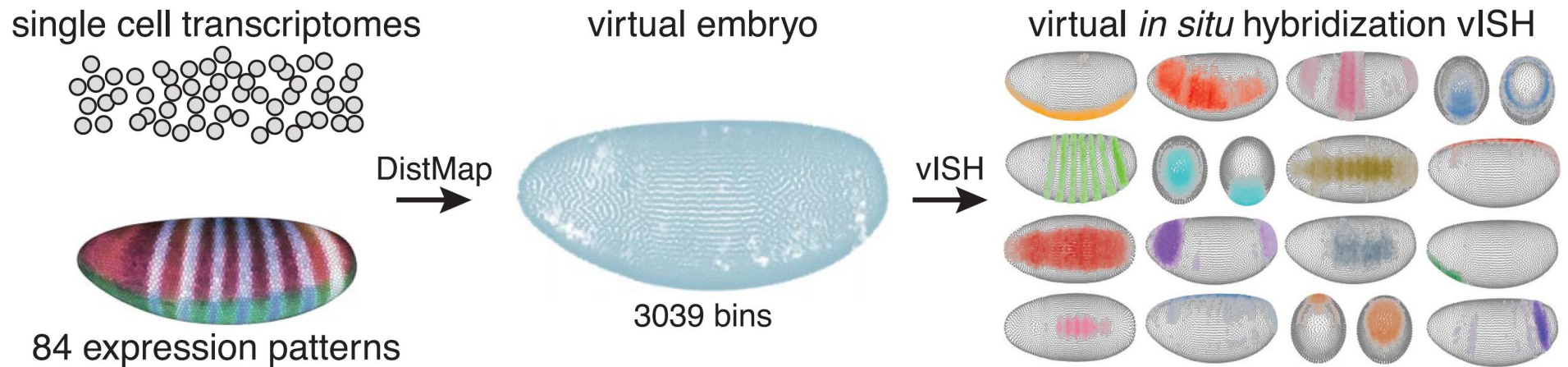
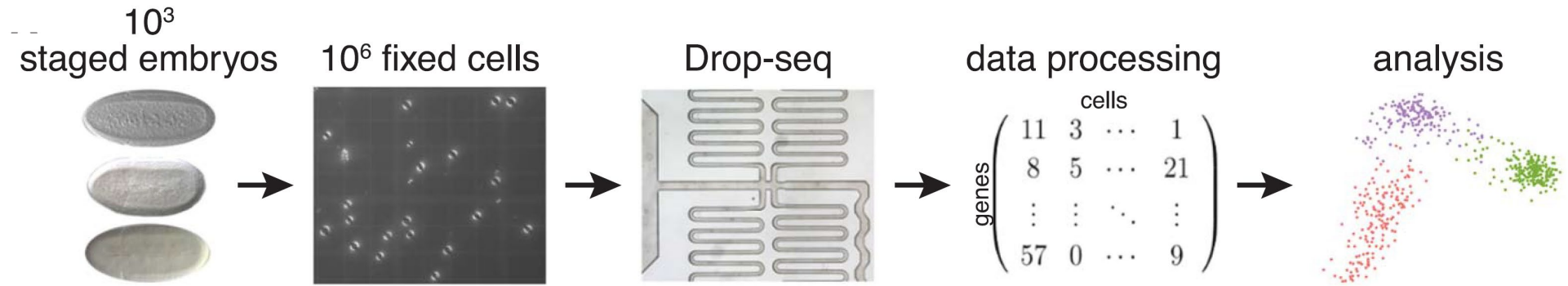
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# Reference-based or *de novo* reconstruction



# DistMap reconstructed *Drosophila* embryo based on ISH references

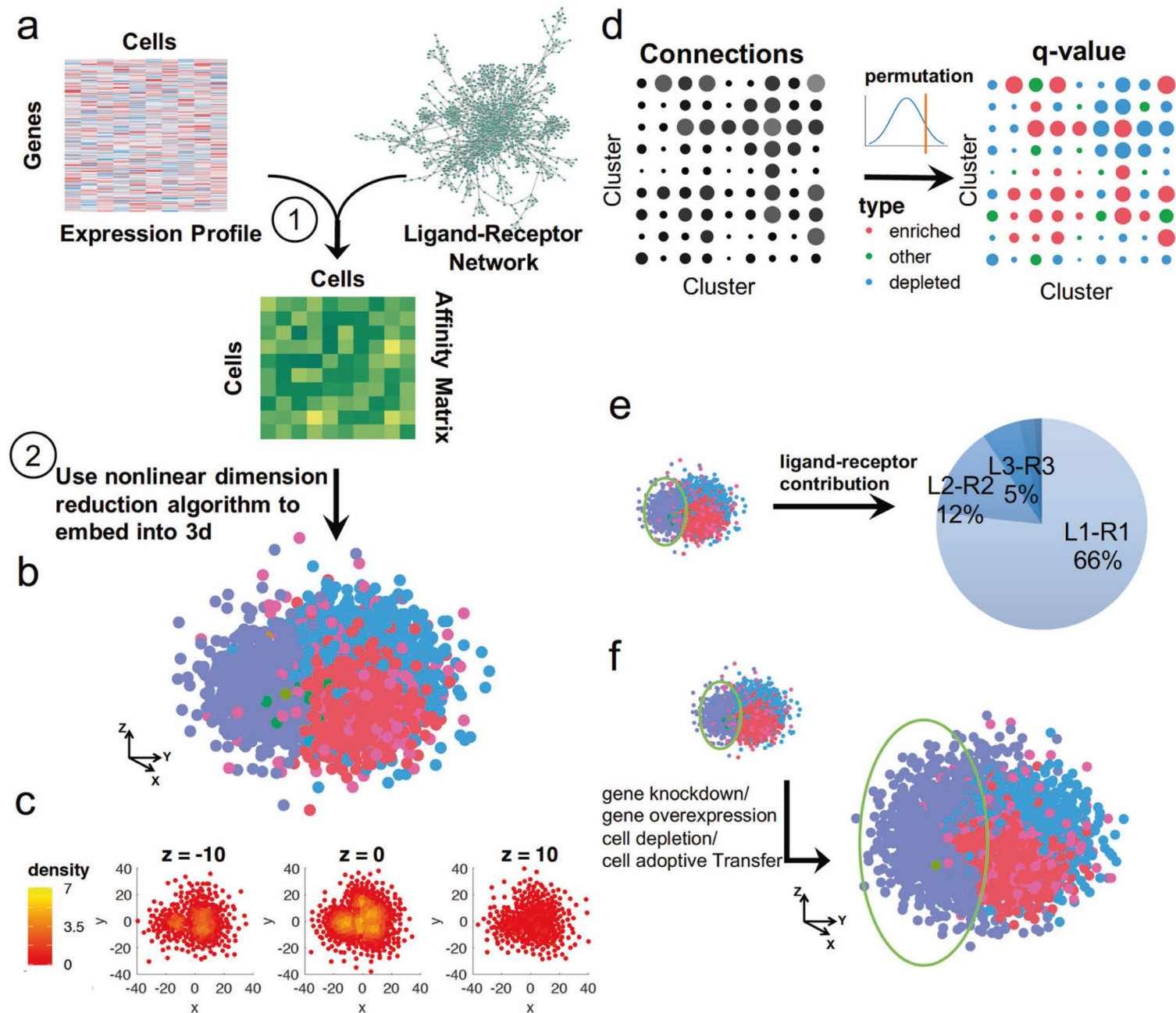
+ Cellular resolution – Require ISH database – Can't be applied to limited samples



vISH: virtual *in situ* hybridization

Computational methods

# CSOmap reconstructed cell organization based on ligand-receptor interactions



# Four paradigms of spatial transcriptomics

Spatial info. source	Microdissection	Imaging-based methods		Spatial mapping	Computation
Key feature	Dissociation	<i>In situ</i> hybridization	<i>In situ</i> sequencing	<i>In vivo</i> capturing	<i>In silico</i> reconstruction
Available methods	Geo-seq (LCM) tomo-seq TIVA NICH-seq ProxmID	smFISH seqFISH MERFISH <b>seqFISH+</b>	Padlock probes BaristaSeq STARmap FISSEQ	ST Slide-seq HDST DBiT-seq	DistMap novoSpaRc <b>CSOmap</b>

# The end of introduction

Any question?

